

# Dietary Fats and Health: Dietary Recommendations in the Context of Scientific Evidence<sup>1</sup>

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## ABSTRACT

Although early studies showed that saturated fat diets with very low levels of PUFAs increase serum cholesterol, whereas other studies showed high serum cholesterol increased the risk of coronary artery disease (CAD), the evidence of dietary saturated fats increasing CAD or causing premature death was weak. Over the years, data revealed that dietary saturated fatty acids (SFAs) are not associated with CAD and other adverse health effects or at worst are weakly associated in some analyses when other contributing factors may be overlooked. Several recent analyses indicate that SFAs, particularly in dairy products and coconut oil, can improve health. The evidence of  $\omega 6$  polyunsaturated fatty acids (PUFAs) promoting inflammation and augmenting many diseases continues to grow, whereas  $\omega 3$  PUFAs seem to counter these adverse effects. The replacement of saturated fats in the diet with carbohydrates, especially sugars, has resulted in increased obesity and its associated health complications. Well-established mechanisms have been proposed for the adverse health effects of some alternative or replacement nutrients, such as simple carbohydrates and PUFAs. The focus on dietary manipulation of serum cholesterol may be moot in view of numerous other factors that increase the risk of heart disease. The adverse health effects that have been associated with saturated fats in the past are most likely due to factors other than SFAs, which are discussed here. This review calls for a rational reevaluation of existing dietary recommendations that focus on minimizing dietary SFAs, for which mechanisms for adverse health effects are lacking. *Adv. Nutr.* 4: 294–302, 2013.

## Introduction

Since the Framingham Heart Study reported that high serum cholesterol was a major risk factor for coronary heart disease (1), there has been an aggressive campaign in the medical community to decrease serum cholesterol. It has been a widely accepted belief that dietary saturated fats and dietary cholesterol cause an increase in serum total cholesterol, as well as LDL-cholesterol (LDL-C)<sup>2</sup> and thereby increase the risk of heart disease if consumed (2). Over the years, it became clear that high levels of LDL circulating in the blood are susceptible to lipid peroxidation, which results in the oxidized LDL being scavenged by macrophages lining certain arteries, particularly around the heart, leading to atherosclerosis (3). Although this mechanism provides a role for high serum LDL-C causing atherosclerosis, evidence of the involvement of saturated fats is lacking, even though it is well established that a diet high in saturated fat increases

serum cholesterol and a diet high in polyunsaturated oil decreases serum cholesterol (4,5). In fact, PUFAs are the components that are oxidized and generate antigenic substances that are recognized by immune cells for clearance of oxidized LDL in atherogenesis (6–8).

Numerous reports and reviews in recent years have begun to call the perceived pernicious effects of dietary saturated fatty acids (SFAs) into question. The purpose of this review is to summarize the scientific understanding as it relates to dietary fats in health and disease, particularly with regard to the innocuous nature of SFAs and the physiological effects that have implicated PUFAs in numerous disorders and diseases. The role of dietary fats in cardiovascular disease (CVD) and many other diseases is complex, yet there is a powerful inertia that has allowed the saturated fat doctrine to endure.

## Dietary fatty acids and serum cholesterol

Dietary fat studies in the mid-20th century stressed the relationship of dietary SFAs and PUFAs to serum cholesterol levels with an aim toward decreasing the likelihood of the development of coronary artery disease (CAD) and premature death (4,5). Once lipoprotein fractions were separated in the blood, it became evident that LDL and VLDL were

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<sup>2</sup> Abbreviations used: apo B, apolipoprotein B-100; CAD, coronary artery disease; CVD, cardiovascular disease; FH, familial hypercholesterolemia; HDL-C, HDL cholesterol; HFCS, high fructose corn syrup; LDL-C, LDL cholesterol; Lp(a), lipoprotein(a); PON1, paraoxonase 1; SFA, saturated fatty acid; SREBP, sterol regulatory element binding protein.

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the carriers of cholesterol that were most closely associated with risk of heart disease (9). Later it was found that the ratio of total serum cholesterol to HDL-C was a better indicator of heart disease risk (10). By the 1990s, the mechanisms by which dietary fats and specific types of fatty acids were regulating serum cholesterol and lipoproteins were beginning to be revealed.

A family of proteins known as sterol regulatory element binding proteins (SREBPs) were discovered in the early 1990s. These proteins move to the nucleus in cholesterol-depleted cells to alter transcription of several genes involved in lipid metabolism (11). When intracellular cholesterol levels are low, SREBP-1 promotes expression of genes for synthesis of cholesterol and LDL receptors that remove cholesterol from the circulation. When intracellular cholesterol levels are high, SREBP-1 is not activated by protease cleavage, and the genes for cholesterol production and LDL receptors are downregulated. SREBP-1 also activates promoters for genes involved in fatty acid synthesis and lipid storage (12). PUFAs, particularly docosahexaenoic acid and others to a lesser extent, regulate expression of the SREBP genes (13,14). Consequently, when PUFAs are present, there is less expression of SREBPs and enzymes for cholesterol synthesis, and the serum cholesterol pool decreases.

There appears to be a number of proteins that bind PUFAs and are involved in regulating gene expression, including a family of G protein-coupled receptors (15), as well as peroxisome proliferator-activated receptors- $\alpha$  and - $\gamma$ , retinoid X receptors, and various other nuclear receptors (16). The liver uses a variety of these receptors or sensors for PUFAs to regulate storage and utilization versus oxidation of PUFAs (17). In this way, PUFAs can stimulate fatty acid oxidation in the liver to minimize their potential for free radical oxidation in the body when their levels are high in the diet. One must keep in mind that this complex array for regulation of expression of a wide range of genes is also subject to an even more complex array of responses to dietary PUFAs and other dietary factors.

Single nucleotide polymorphisms in genes for many of the above factors, as well as in genes for several apolipoproteins, TNFs, glutathione peroxidases, and other proteins result in a wide range of individual responses to dietary constituents. The consequences of such genetic variation can be either little change or very large changes in serum lipids and lipoproteins in response to diet, depending on an individual's genetic make-up (18). However, one should not lose sight of the fact that levels of many other proteins are being altered in the process, which can give rise to a wide array of physiological responses that influence susceptibility to many unhealthy conditions, such as CVD and cancer.

Short-chain SFAs, such as those in dairy fat and coconut oil, can also influence gene expression via interactions with various G protein-coupled receptors that are linked to several hormonal responses, including insulin and leptin, that regulate overall energy metabolism in the body (19). It is clear that there are numerous sensors that respond to dietary PUFAs and short- or medium-chain SFAs (20).

## Genetic factors

Brown and Goldstein (21) received the Nobel Prize in Physiology or Medicine in 1985 for their work on genetic defects in LDL receptors of people with familial hypercholesterolemia (FH). They identified several mutations that produce nonfunctional LDL receptors, resulting in death from atherosclerosis and heart disease at an early age. Individuals with FH have serum LDL-C in excess of 300 mg/dL (or 8 mmol/L), although LDL-C may be as high as 650 mg/dL (17 mmol/L) in homozygous individuals. Goldstein and Brown (22) also identified several genes that code for other proteins involved in cholesterol transport and metabolism, such as apolipoprotein B-100 (apo B), which is a component of LDL that binds to LDL receptors. There are other proteins involved in LDL synthesis, transport, and clearance that can result in a genetic predisposition to increased serum LDL cholesterol and FH (23–25).

In the early 1990s, it was discovered that men with CVD tended to have smaller HDL particles than healthy controls (26). It was later found that LDL particle size was also significantly smaller in men with CAD than in case-matched controls (27), although another study showed the ratio of total serum cholesterol to HDL-C was a better predictor of CAD risk than LDL particle size (28). A prospective, population-based cohort study also found an increased risk of CAD in middle-aged men with smaller, dense LDL particles than in men with larger LDL particles, although the relationship did not show a linear dependence on particle size (29). It later became evident that LDL particle size was influenced by several factors and was not necessarily a useful predictor of heart disease risk; the nature of LDL is influenced by both dietary and genetic factors (30).

Lipoprotein (a) [Lp(a)] is a complex lipoprotein that has several properties in common with LDL. Like LDL and VLDL, Lp(a) contains apo B, but also contains highly variable forms of apolipoprotein(a) that strongly influence its atherogenicity and propensity to promote heart disease (31). The wide array of apolipoprotein(a) isoforms present in the human population may have caused some confusion regarding the role of Lp(a) in atherogenesis and CVD. The association of apo B with oxidized phospholipids was found to be dependent on Lp(a) (32). The presence of oxidized phospholipids and Lp(a) tend to be proinflammatory and promote atherogenesis.

Small, dense LDL particles rarely occur as an isolated condition, but are often associated with a specific phenotype that is characterized by hypertriglyceridemia, low HDL-C, abdominal obesity, insulin resistance, and other metabolic irregularities that lead to endothelial dysfunction and susceptibility to thrombosis (33). Small, dense LDL is also more susceptible to lipid peroxidation due to changes in the lipid composition, making it more atherogenic (34). LDL particles from the atherogenic phenotype contain less cholesterol and phospholipid, but more triglyceride. This phenotype is generally referred to as phenotype B and is characterized by elevated levels of apo B, which is found in LDL and VLDL (35).

There have been a host of proteins linked to lipoprotein metabolism and transport and a wide range of genetic variations identified that result in alterations of those proteins. Many are associated with HDL and larger HDL particle size, which is consistently associated with a decreased risk of CAD (36). HDL is important in reverse-cholesterol transport, bringing cholesterol from arterial deposits to the liver for processing, where it is converted to useful metabolites and eventually cleared from the body via bile secretions. A family of lipoprotein lipases, including hepatic lipase and endothelial lipase, are intimately involved in HDL metabolism. Endothelial lipase is upregulated during inflammation, a condition that increases LDL oxidation and atherogenesis (37). Genetic variation in apolipoprotein A-I, a major protein component of HDL, can result in larger but less stable HDL particles and decreased levels of circulating HDL (38). Cholesteryl ester transfer protein is generally considered to be protective, although this protein may transfer lipids from HDL to other lipoproteins that result in a less desirable serum lipid profile (39). HDL is emerging as a fascinating lipoprotein with a complex array of functions that involve both protein and lipid components. HDL has been found to influence immune function, vascular inflammation, glucose metabolism, and platelet function as well as other physiological phenomena unrelated to CVD (40).

Paraoxonase 1 (PON1) is another protein associated with HDL that exhibits esterase and lactonase enzyme activity, including metabolism of toxic organophosphorus pesticides and oxidized lipids in oxidized LDL particles. The levels of PON1 activity varies tremendously among humans, which depends to a large degree on genetic variation. However, environmental factors, such as dietary antioxidant consumption, alcohol consumption, and certain drugs can also influence PON1 activity (41). Dietary olive oil can increase levels of serum PON1 in some individuals, which is genotype dependent (42), whereas MUFAs and PUFAs can inhibit PON1 enzymatic activity (43). SFAs (palmitic and myristic) had virtually no effect on PON1 enzymatic activity. A recent study found that HDL isolated from patients with CAD lacks endothelial anti-inflammatory properties, has lower PON1 enzyme activity, and does not promote endothelial nitric oxide production (44), all of which are most likely tied to genetic rather than dietary factors.

### **Fatty acids involved in atherogenesis and CVD**

Linoleic acid makes LDL more susceptible to lipid peroxidation and subsequent deposition of the oxidized LDL in macrophages lining the arteries (45). Several lipid peroxidation products have been shown to trigger transformation of circulating monocytes to macrophages that line the arteries and ultimately become foam cells (46,47). Lipid peroxidation products also signal cells in the arterial intima to encapsulate foam cells by surrounding them with extracellular matrix proteins and eventually calcify the matrix (48). It would stand to reason that a greater abundance of PUFAs, relative to SFAs and MUFAs, during conditions of oxidative stress would provoke atherogenesis. The fibrous cap that is

formed over fatty deposits makes them inaccessible to apolipoproteins such as apolipoprotein A-I or E, which are components of HDL, the lipoprotein that removes cholesterol from these deposits (49). The protein cap is characteristic of advanced atherosclerotic plaque and erosion of this protective cap by extracellular metalloproteases can release collagen and collagen-like fragments that trigger blood platelets to initiate a blood clot, which results in myocardial infarction or stroke (3).

Because saturated fats are not susceptible to lipid peroxidation, they have not been found to be involved in these mechanisms. This begs the question of how dietary polyunsaturated oils seem to lower the risk of CAD, even though many studies have shown no such effect. One important consideration is that foods that are considered sources of predominantly saturated fats, such as meats, are often cooked at high temperatures, which can induce lipid peroxidation in the minor amounts of PUFAs present in those animal products (50–52). Oxidative stress and lipid peroxidation products are known to promote heart disease, cancer, and several other chronic diseases (53,54). High-temperature cooking can also oxidize carbohydrates, producing a range of toxic oxidation products that promote oxidative stress, type 2 diabetes, and CVD (55). The preparation and cooking methods used for foods that are traditionally classified as saturated fat foods may be producing substances from PUFAs and carbohydrates in those foods that are promoting disease.

Human food preferences tend to favor foods with both fats and sugar (56), which complicates any attempts to correlate saturated fats with disease. Sugars readily undergo oxidation, with fructose generally getting oxidized many times faster than glucose, whereas sucrose is relatively resistant to oxidation (57). The oxidation products of these monosaccharides include glyoxal, methylglyoxal, and formaldehyde. Methylglyoxal has been shown to promote endothelial dysfunction as well as hypercholesterolemia in rats (58). Methylglyoxal is also associated with increased atherosclerosis and hypertension in humans (59). Formaldehyde and methylglyoxal have been implicated in endothelial injury, oxidative stress, and angiopathy (60).

Many clinical studies show that there are fewer coronary events when polyunsaturated oils replace saturated fats in the diet (61). However, a recent meta-analysis found that interventions using mixed  $\omega$ 3 and  $\omega$ 6 PUFAs resulted in a significant (22%) decrease in CAD events compared with control diets with fewer PUFAs. However, interventions that used  $\omega$ 6 polyunsaturated oils with no  $\omega$ 3 PUFAs showed ~16% more cardiovascular events compared with the control diets, although the increased number was not statistically significant (62). It would seem that even moderate amounts of  $\omega$ 3 PUFAs in the diet result in attenuation of inflammatory responses that are reflected in the significant reduction in coronary events observed with increasing dietary PUFAs. Of the common vegetable oils, soy oil contains ~7%  $\omega$ 3 PUFAs and canola oil as much as 10%  $\omega$ 3 PUFAs, whereas corn, safflower, and sunflower oils generally contain

<1%  $\omega$ 3 PUFAs (63). Another systematic review found insufficient evidence to support an association (positive or negative) between CAD and several dietary factors, including SFAs or PUFAs,  $\alpha$ -linolenic acid, total fat, meat, eggs, and milk (64).

### Lipid peroxidation and inflammation

Lipid peroxidation is invoked as a mechanism for numerous adverse health effects, such as aging, cancer, atherosclerosis, and tissue necrosis. The greater in vivo susceptibility of  $\omega$ 6 PUFAs relative to the  $\omega$ 3 PUFAs, has placed the spotlight on these fatty acids as contributing to or exacerbating many ailments (68). The metabolism of arachidonic acid to bioactive eicosanoids is responsible for many of the biological processes that lead to inflammation. Indeed, steroidal and nonsteroidal anti-inflammatory drugs suppress inflammation by blocking the release of arachidonic acid from membranes or its subsequent metabolism to eicosanoids.

Studies of inflammation in rats have found that dietary manipulation of relative amounts of  $\omega$ 6 PUFA precursors can have profound effects on the degree of inflammation. Predominantly SFAs in the diet result in far less inflammation than diets with either  $\omega$ 3 (69) or  $\omega$ 6 PUFAs (70). Several studies have shown that dietary supplementation with  $\omega$ 3 PUFAs can reduce inflammation and make patients less dependent on drug therapy to manage the pain and stiffness of arthritis (71–73). Patients should be advised to minimize their intake of  $\omega$ 6 oils when attempting  $\omega$ 3 supplementation as a therapeutic approach to reduce the inflammation of arthritis and other inflammatory syndromes (74,75). Small amounts of  $\omega$ 3 supplements in a sea of dietary  $\omega$ 6 oils would have relatively little chance of changing the course of an inflammatory response. Because dietary saturated fats do not promote inflammation, it may be wiser to minimize  $\omega$ 6 PUFAs and consume more SFAs to reduce various types of inflammation; most sources of MUFAs contain significant amounts of PUFAs as well. There have been few scientific studies along these lines because of the misguided concern that saturated fats, even those from vegetable sources such as palm and coconut oil, would be detrimental to one's health.

The efficacy of  $\omega$ 3 supplements for inflammatory syndromes other than rheumatoid arthritis are less persuasive, although study designs are questioned regarding whether patients are advised to reduce their  $\omega$ 6 fatty acid intake (76). Fish oil supplements improved pulmonary function in some asthmatics (responders) but not in others (nonresponders). A relatively high ratio (10:1) of dietary  $\omega$ 6 to  $\omega$ 3 PUFAs resulted in diminished respiratory function in methacholine-provoked asthmatics, whereas a lower ratio (2:1) produced significant improvement in >40% of the study participants (77). A study in Japan showed beneficial effects of  $\omega$ 3 supplements in asthmatic children in a controlled hospital ward environment (78). A comparison of dietary saturated fats with polyunsaturated oils was not found in the literature for asthma studies. Such an approach would be logical for this life-threatening condition, in view of the

benign nature of saturated fats and the fact that carbohydrates, especially sugars, may actually be augmenting the incidence of asthma (79).

### Are low-fat, low-saturated fat diets healthier?

Studies with laboratory animals have shown that high-fat diets promote chemically induced cancers (80,81). A study of chemically induced mammary tumors in rats found that  $\omega$ 6 PUFAs promoted tumor proliferation, whereas saturated fats or  $\omega$ 3 PUFAs did not promote tumors as much or even suppressed tumors, depending on what one uses as a reference (82,83). Although 1 review and meta-analysis found that linoleic acid, the predominant  $\omega$ 6 fatty acid in vegetable oils, is not a risk factor for breast, colorectal, and prostate cancers in humans (84), there is evidence to the contrary that high intake of  $\omega$ 6 relative to  $\omega$ 3 PUFAs increases cancer risks (85–87). There are multiple processes by which  $\omega$ 6 fatty acids can promote carcinogenesis; production of bioactive eicosanoids from arachidonic acid is 1 mechanism (88,89). Nonsteroidal anti-inflammatory drugs as well as cyclooxygenase-2 inhibitors can suppress tumors by inhibiting production of prostaglandins, particularly those of the  $\omega$ 6 variety (90). Lipid peroxides are also known to promote chemically induced tumors (91), and PUFAs are highly susceptible to lipid peroxidation.

Investigators often seem to have a particular bias against saturated fats. One report showed that red meat alone was not significantly associated with colorectal cancer, although there was some increase in colorectal cancers with higher red meat intake [HR = 1.17 for highest vs. lowest intakes (95% CI = 0.92–1.49,  $P_{\text{trend}} = 0.08$ )]. Processed meats were significantly associated [HR = 1.42 (95% CI = 1.09–1.86,  $P_{\text{trend}} = 0.02$ )]. The authors then combined the data for red meat and processed meat to give a significant association and concluded that red and processed meat are positively associated with colorectal cancer (92). When specific types of meat were analyzed, significant risk was associated with pork [HR = 1.18 (95% CI = 0.95–1.48,  $P_{\text{trend}} = 0.02$ )] and lamb [HR = 1.22 (95% CI = 0.96–1.55,  $P_{\text{trend}} = 0.03$ )], but not with beef/or veal [HR = 1.03 (95% CI = 0.86–1.24,  $P_{\text{trend}} = 0.76$ )]. It is interesting to note that in 1 study, beef had a much lower ratio of PUFAs to SFAs than pork, but nearly the same ratio of PUFAs to SFAs as sheep (93). The ratio of MUFAs to SFAs in beef also varies, as it does in most meats, with the ratio ranging from  $\sim$ 0.8 to 1.8, depending on breed and feeding practices (94).

Nitrite used in the preservation of many processed meats is known to form a carcinogen with secondary amines under acidic conditions that would prevail in the stomach (95). Others have found no association of red meat and only a very weak association of processed meat with breast cancer (96) and prostate cancer (97). Most studies find no differences in cancer risk with different types of fat, but do find associations with high levels of fat in the diet (81).

A recent meta-analysis (98) reviewed 20 studies with >1 million subjects and found that red meat was not associated with CAD events [RR = 1.00 (95% CI = 0.81–1.23,  $P_{\text{trend}} = 0.36$ )].

In contrast, processed meats were associated with increased incidence of CAD [RR = 1.42 (95% CI = 1.07–1.89,  $P_{\text{trend}} = 0.04$ )]. This indicates that saturated fat per se is not increasing CAD events, but other factors are, such as preservatives used in processed meats or other dietary substances that are being consumed in conjunction with processed meats. It is important to keep in mind that meats generally contain as much MUFA as SFA. Others are beginning to challenge the saturated fat hypothesis with closer analyses of past studies (99–103).

Campaigns were waged against tropical oils (palm and coconut oils) in the early 1980s because of their high levels of SFAs, even though palm oil contains about as much MUFAs acids as SFAs and has an ample amount of PUFAs to keep serum cholesterol low. In fact, 2 studies showed that the higher ratio of SFAs to MUFAs in palm oil (1.1:1) compared with olive oil (0.22:1) had no effect on serum lipids in healthy volunteers (104,105). Palm oil and olive oil have similar amounts (~10%) of PUFAs. SFAs in coconut oil increase serum HDL-C more than LDL-C to give a more favorable lipid profile relative to dietary carbohydrates (10). Claims that tropical oils with a high SFA content increase the risk of CAD lack clear scientific evidence to that effect. Indeed, countries with high intake of tropical oils have some of the lowest rates of heart disease in the world (106).

Many of the shorter chain fatty acids found in milk fat and coconut oil have beneficial health effects. The shorter chain SFA in milk (C4–C12) are not only metabolized rapidly for energy in infants, but have been found to have important antiviral, antimicrobial, antitumor, and immune response functions (107). Lauric acid, which is present in milk and the most abundant fatty acid in coconut oil, is effective in preventing tooth decay and plaque buildup (108). Diets rich in coconut oils have also been shown to lower other risk factors for CAD, such as tissue plasminogen activator antigen and Lp(a) (109). The medium-chain SFAs in coconut oil and butterfat (milk) increase total serum cholesterol, but their positive effects on HDL-C are protective in many ways. There is also evidence that proteins, fats, and calcium in milk are beneficial in lowering blood pressure, inflammation, and the risk of type 2 diabetes (110,111). Indeed, these constituents of milk have clear beneficial effects against metabolic syndrome, which is a major factor in promoting heart disease, as well as premature death from a variety of causes (112).

There has been a spate of recent publications in the biomedical literature that question the negative perception that dairy fats are bad for health. One meta-analysis showed that participants in prospective studies with the highest consumption of dairy products had a lower RR for all-cause mortality as well as for CAD, stroke, and diabetes compared with the lowest intake of dairy products (113). Many of the studies included in the analysis started before low-fat milk was available on the market. Another review arrived at the same conclusion that consumption of dairy products is not associated with higher risk of CVD (100). Although prospective cohort studies often find a significant reduction in

the incidence of CAD with a larger ratio of PUFAs to SFAs in the diet (114), there are often many other factors related to overall health that correlate with the unsaturated to SFA ratio, such as exercise, a healthier lifestyle, and more fiber and less sugar in the diet.

### **Less fat generally means more carbohydrate**

It should not be surprising that substitution of carbohydrates (starches) for saturated fats in the diet has relatively little effect on serum lipids. Excess carbohydrates are converted to fats for efficient energy storage, and the human body synthesizes primarily SFAs from excess carbohydrates, although MUFAs are also formed. Consequently, from a physiological viewpoint, there is no reason to believe that replacing fat in the diet with carbohydrate at a constant caloric intake will improve the serum lipid profile significantly. Indeed, a low-fat, high-carbohydrate diet causes an increase in serum triglycerides and small, dense LDL particles (115), which are more strongly associated with CAD than serum total cholesterol or LDL-C. When dietary fat is replaced by carbohydrate without changing the fatty acid composition of the fat, there is no change in LDL-C or HDL-C, but there is an increase in serum triglycerides (116). However, if there is a higher percentage of PUFAs and lower SFAs in a low-fat diet, serum total cholesterol and LDL-C will decrease (117).

Young children who consumed more fruit juice than their peers were shorter in stature and had greater BMI than their peers who drank less fruit juice (118). A trend of increased fruit juice consumption by infants and children in recent years has coincided with a decrease in milk consumption (119). The rates of childhood obesity have skyrocketed since the introduction of low-fat milk, although high fructose corn syrup (HFCS) became omnipresent in foods at the same time and is more strongly associated with obesity than dietary fat (120,121). As stated previously, the short-chain SFAs in milk provide valuable antibacterial and antiviral activities, which would result in healthier children. The short-chain SFAs found in milk act as signaling agents in the immune system (122). Infections in children also correlated with higher levels of atherogenic oxidized LDL, as well as lower levels of HDL (123). It is possible that oxidized LDL and low HDL impart increased susceptibility to infection, although the combination of infections and an adverse serum lipid profile may both result from an undesirable diet, i.e., more sugar and fewer healthy fats.

Food processors generally add large amounts of sugar to fat-free or low-fat foods to make them more palatable to consumers. Fructose is 1 dietary constituent that is consistently found to have adverse health consequences, and the larger the proportion of fructose is in the diet, the more formidable the effect. The adverse effects of fructose that have been documented include increased serum triglycerides, particularly in men (124,125); increased serum uric acid, which is associated with gout and hypertension (126); increased lipid peroxidation (57) and increased oxidation of LDL (127); increased oxidative stress in animal models (128); greater risk of the development of metabolic syndrome,

including obesity, insulin resistance, hypertension, and CVD risk (129,130); increased nonalcoholic fatty liver disease (131); and increased systemic inflammation and associated renal disease (132).

There are clearly many established physiological mechanisms by which fructose increases CVD and several other diseases. Whether the source of dietary fructose is sucrose or HFCS would seem irrelevant, although sucrose is 50% fructose, whereas the most common dietary source of HFCS (soft drinks) is generally 55% fructose and ~43% glucose. Solutions of fructose are also highly susceptible to autoxidation, producing a host of toxic products (57), whereas sucrose is highly resistant to oxidation. The toxic products from fructose oxidation include formaldehyde and  $\alpha$ -dicarbonyls. Although saturated fats have been implicated in many of the adverse health effects attributed to fructose, there is no scientific evidence to support a role for saturated fats in the physiological mechanisms. On the other hand, plausible mechanisms are proposed for all of the unhealthy conditions promoted by high fructose intake mentioned earlier.

It turns out that a high level of fructose in the diet increases plasma triglycerides, which leads to not only increased levels of VLDL and small, dense LDL particles, but increased levels of oxidized LDL, insulin resistance, and other metabolic consequences linked to metabolic syndrome and dyslipidemia (133). The mechanisms by which fructose promotes inflammation and elevated levels of uric acid and several cytokines have been reviewed (132).

## Conclusions

Saturated fats are benign with regard to inflammatory effects, as are the MUFAs. The meager effect that saturated fats have on serum cholesterol levels when modest but adequate amounts of polyunsaturated oils are included in the diet, and the lack of any clear evidence that saturated fats are promoting any of the conditions that can be attributed to PUFA makes one wonder how saturated fats got such a bad reputation in the health literature. The influence of dietary fats on serum cholesterol has been overstated, and a physiological mechanism for saturated fats causing heart disease is still missing.

Various aldehydes produced in the oxidation of PUFAs, as well as sugars, are known to initiate or augment several diseases, such as cancer, inflammation, asthma, type 2 diabetes, atherosclerosis, and endothelial dysfunction. Saturated fats per se may not be responsible for many of the adverse health effects with which they have been associated; instead, oxidation of PUFAs in those foods may be the cause of any associations that have been found. Consequently, the dietary recommendations to restrict saturated fats in the diet should be revised to reflect differences in handling before consumption, e.g., dairy fats are generally not heated to high temperatures. It is time to reevaluate the dietary recommendations that focus on lowering serum cholesterol and to use a more holistic approach to dietary policy.

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## Literature Cited

1. Kannel WB, Dawber TR, Kagan A, Revotskie N, Stokes J. Factors of risk in the development of coronary heart disease: six-year follow-up experience. *Ann Intern Med.* 1961;55:33–50.
2. Steinberg D. Thematic review series: the pathogenesis of atherosclerosis. An interpretive history of the cholesterol controversy: part II: the early evidence linking hypercholesterolemia to coronary disease in humans. *J Lipid Res.* 2005;46:179–90.
3. Libby P. Changing concepts of atherogenesis. *J Intern Med.* 2000;247:349–58.
4. Hegsted DM, McGandy RB, Myers ML, Stare FJ. Quantitative effects of dietary fat on serum cholesterol in man. *Am J Clin Nutr.* 1965;17:281–95.
5. Keys A, Anderson JT, Grande F. Prediction of serum-cholesterol responses of man to changes in fats in the diet. *Lancet.* 1957;273:959–66.
6. Morel DW, Hessler JR, Chisolm GM. Low density lipoprotein cytotoxicity induced by free radical peroxidation of lipid. *J Lipid Res.* 1983;24:1070–6.
7. Steinbrecher UP, Parthasarathy S, Leake DS, Witztum JL, Steinberg D. Modification of low density lipoprotein by endothelial cells involves lipid peroxidation and degradation of low density lipoprotein phospholipids. *Proc Natl Acad Sci U S A.* 1984;81:3883–7.
8. Navab M, Ananthramaiiah GM, Reddy ST, Van Lenten BJ, Ansell BJ, Fonarow GC, Vahabzadeh K, Hama S, Hough G, Kamranpour N, et al. The oxidation hypothesis of atherogenesis: the role of oxidized phospholipids and HDL. *J Lipid Res.* 2004;45:993–1007.
9. Steinberg D. Thematic review series: the pathogenesis of atherosclerosis: an interpretive history of the cholesterol controversy, part III: mechanistically defining the role of hyperlipidemia. *J Lipid Res.* 2005;46:2037–51.
10. Mensink RP, Zock PL, Kester AD, Katan MB. Effects of dietary fatty acids and carbohydrates on the ratio of serum total to HDL cholesterol and on serum lipids and apolipoproteins: a meta-analysis of 60 controlled trials. *Am J Clin Nutr.* 2003;77:1146–55.
11. Brown MS, Goldstein JL. The SREBP pathway: regulation of cholesterol metabolism by proteolysis of a membrane-bound transcription factor. *Cell.* 1997;89:331–40.
12. Lopez JM, Bennett MK, Sanchez HB, Rosenfeld JM, Osborne TF. Sterol regulation of acetyl coenzyme A carboxylase: a mechanism for coordinate control of cellular lipid. *Proc Natl Acad Sci U S A.* 1996;93:1049–53.
13. Jump DB. N-3 polyunsaturated fatty acid regulation of hepatic gene transcription. *Curr Opin Lipidol.* 2008;19:242–7.
14. Xu J, Nakamura MT, Cho HP, Clarke SD. Sterol regulatory element binding protein-1 expression is suppressed by dietary polyunsaturated fatty acids. A mechanism for the coordinate suppression of lipogenic genes by polyunsaturated fats. *J Biol Chem.* 1999;274:23577–83.
15. Covington DK, Briscoe CA, Brown AJ, Jayawickreme CK. The G-protein-coupled receptor 40 family (GPR40–GPR43) and its role in nutrient sensing. *Biochem Soc Trans.* 2006;34:770–3.
16. Bordoni A, Di Nunzio M, Danesi F, Biagi PL. Polyunsaturated fatty acids: from diet to binding to ppars and other nuclear receptors. *Genes Nutr.* 2006;1:95–106.
17. Clarke SD. The multi-dimensional regulation of gene expression by fatty acids: polyunsaturated fats as nutrient sensors. *Curr Opin Lipidol.* 2004;15:13–8.
18. Curti ML, Jacob P, Borges MC, Rogero MM, Ferreira SR. Studies of gene variants related to inflammation, oxidative stress, dyslipidemia, and obesity: implications for a nutrigenetic approach. *J Obes* 2011; 2011:497401.
19. Ichimura A, Hirasawa A, Hara T, Tsujimoto G. Free fatty acid receptors act as nutrient sensors to regulate energy homeostasis. *Prostaglandins Other Lipid Mediat.* 2009;89:82–8.

20. Hara T, Hirasawa A, Ichimura A, Kimura J, Tsujimoto G. Free fatty acid receptors FFAR1 and GPR120 as novel therapeutic targets for metabolic disorders. *J Pharm Sci*. 2011;100:3594–601.
21. Brown MS, Goldstein JL. A receptor-mediated pathway for cholesterol homeostasis. *Science*. 1986;232:34–47.
22. Goldstein JL, Brown MS. Molecular medicine. The cholesterol quartet. *Science*. 2001;292:1310–2.
23. Mishra SK, Watkins SC, Traub LM. The autosomal recessive hypercholesterolemia (ARH) protein interfaces directly with the clathrin-coat machinery. *Proc Natl Acad Sci U S A*. 2002;99:16099–104.
24. Liyanage KE, Burnett JR, Hooper AJ, van Bockxmeer FM. Familial hypercholesterolemia: epidemiology, Neolithic origins and modern geographic distribution. *Crit Rev Clin Lab Sci*. 2011;48:1–18.
25. Fahed AC, Nemer GM. Familial hypercholesterolemia: the lipids or the genes? *Nutr Metab (Lond)*. 2011;8:23.
26. Cheung MC, Brown BG, Wolf AC, Albers JJ. Altered particle size distribution of apolipoprotein A-I-containing lipoproteins in subjects with coronary artery disease. *J Lipid Res*. 1991;32:383–94.
27. Stampfer MJ, Krauss RM, Ma J, Blanche PJ, Holl LG, Sacks FM, Hennekens CH. A prospective study of triglyceride level, low-density lipoprotein particle diameter, and risk of myocardial infarction. *JAMA*. 1996;276:882–8.
28. Gardner CD, Fortmann SP, Krauss RM. Association of small low-density lipoprotein particles with the incidence of coronary artery disease in men and women. *JAMA*. 1996;276:875–81.
29. Lamarche B, St-Pierre AC, Ruel IL, Cantin B, Dagenais GR, Despres JP. A prospective, population-based study of low density lipoprotein particle size as a risk factor for ischemic heart disease in men. *Can J Cardiol*. 2001;17:859–65.
30. Bossé Y, Perusse L, Vohl MC. Genetics of LDL particle heterogeneity: from genetic epidemiology to DNA-based variations. *J Lipid Res*. 2004;45:1008–26.
31. Anuurad E, Enkhmaa B, Berglund L. Enigmatic role of lipoprotein(a) in cardiovascular disease. *Clin Transl Sci*. 2010;3:327–32.
32. Tsimikas S, Clopton P, Brilakis ES, Marcovina SM, Khera A, Miller ER, de Lemos JA, Witztum JL. Relationship of oxidized phospholipids on apolipoprotein B-100 particles to race/ethnicity, apolipoprotein(a) isoform size, and cardiovascular risk factors: results from the Dallas Heart Study. *Circulation*. 2009;119:1711–9.
33. Lamarche B, Lemieux I, Despres JP. The small, dense LDL phenotype and the risk of coronary heart disease: epidemiology, patho-physiology and therapeutic aspects. *Diabetes Metab*. 1999;25:199–211.
34. Ohmura H, Mokuno H, Sawano M, Hatsumi C, Mitsugi Y, Watanabe Y, Daida H, Yamaguchi H. Lipid compositional differences of small, dense low-density lipoprotein particle influence its oxidative susceptibility: possible implication of increased risk of coronary artery disease in subjects with phenotype B. *Metabolism*. 2002;51:1081–7.
35. Carmena R, Duriez P, Fruchart JC. Atherogenic lipoprotein particles in atherosclerosis. *Circulation*. 2004;109 23 Suppl 1: III2–7.
36. Chatterjee C, Sparks DL. Hepatic lipase, high density lipoproteins, and hypertriglyceridemia. *Am J Pathol*. 2011;178:1429–33.
37. Yasuda T, Ishida T, Rader DJ. Update on the role of endothelial lipase in high-density lipoprotein metabolism, reverse cholesterol transport, and atherosclerosis. *Circ J*. 2010;74:2263–70.
38. Kono M, Tanaka T, Tanaka M, Vedhachalam C, Chetty PS, Nguyen D, Dhanasekaran P, Lund-Katz S, Phillips MC, Saito H. Disruption of the C-terminal helix by single amino acid deletion is directly responsible for impaired cholesterol efflux ability of apolipoprotein A-I Nichinan. *J Lipid Res*. 2010;51:809–18.
39. Kappelle PJ, van Tol A, Wolffenbuttel BH, Dullaart RP. Cholesteryl ester transfer protein inhibition in cardiovascular risk management: ongoing trials will end the confusion. *Cardiovasc Ther*. 2011;29:e89–99.
40. Gordon SM, Hofmann S, Askew DS, Davidson WS. High density lipoprotein: it's not just about lipid transport anymore. *Trends Endocrinol Metab*. 2011;22:9–15.
41. Costa LG, Giordano G, Furlong CE. Pharmacological and dietary modulators of paraoxonase 1 (PON1) activity and expression: the hunt goes on. *Biochem Pharmacol*. 2011;81:337–44.
42. Tomás M, Senti M, Elosua R, Vila J, Sala J, Masia R, Marrugat J. Interaction between the Gln-Arg 192 variants of the paraoxonase gene and oleic acid intake as a determinant of high-density lipoprotein cholesterol and paraoxonase activity. *Eur J Pharmacol*. 2001;432: 121–8.
43. Nguyen SD, Sok DE. Preferential inhibition of paraoxonase activity of human paraoxonase 1 by negatively charged lipids. *J Lipid Res*. 2004; 45:2211–20.
44. Besler C, Heinrich K, Rohrer L, Doerries C, Riwanto M, Shih DM, Chroni A, Yonekawa K, Stein S. Mechanisms underlying adverse effects of HDL on eNOS-activating pathways in patients with coronary artery disease. *J Clin Invest*. 2011;121:2693–708.
45. Reaven P, Parthasarathy S, Grasse BJ, Miller E, Steinberg D, Witztum JL. Effects of oleate-rich and linoleate-rich diets on the susceptibility of low density lipoprotein to oxidative modification in mildly hypercholesterolemic subjects. *J Clin Invest*. 1993;91:668–76.
46. Subbanagounder G, Leitinger N, Schwenke DC, Wong JW, Lee H, Rizza C, Watson AD, Faull KF, Fogelman AM, Berliner JA. Determinants of bioactivity of oxidized phospholipids. Specific oxidized fatty acyl groups at the sn-2 position. *Arterioscler Thromb Vasc Biol*. 2000; 20:2248–54.
47. Thijssen MA, Mensink RP. Fatty acids and atherosclerosis risk. In: Eckardstein AV, editor. *Atherosclerosis: diet and drugs*. Berlin-Heidelberg, Germany: Springer-Verlag, 2005:165–94.
48. Abedin M, Tintut Y, Demer LL. Vascular calcification: mechanisms and clinical ramifications. *Arterioscler Thromb Vasc Biol*. 2004;24: 1161–70.
49. Demer LL, Tintut Y. Mineral exploration: search for the mechanism of vascular calcification and beyond: the 2003 Jeffrey M. Hoeg Award lecture. *Arterioscler Thromb Vasc Biol*. 2003;23:1739–43.
50. Gatellier P, Sante-Lhoutellier V, Portanguen S, Kondjoyan A. Use of meat fluorescence emission as a marker of oxidation promoted by cooking. *Meat Sci*. 2009;83:651–6.
51. Conchillo A, Ansorena D, Astiasaran I. Combined effect of cooking (grilling and roasting) and chilling storage (with and without air) on lipid and cholesterol oxidation in chicken breast. *J Food Prot*. 2003;66:840–6.
52. Brown ED, Morris VC, Rhodes DG, Sinha R, Levander OA. Urinary malondialdehyde-equivalents during ingestion of meat cooked at high or low temperatures. *Lipids*. 1995;30:1053–6.
53. Cejas P, Casado E, Belda-Iniesta C, De Castro J, Espinosa E, Redondo A, Sereno M, Garcia-Cabezas MA, Vara JA. Implications of oxidative stress and cell membrane lipid peroxidation in human cancer (Spain). *Cancer Causes Control*. 2004;15:707–19.
54. Lankin VZ, Lisina MO, Arzamastseva NE, Konovalova GG, Nedosugova IV, Kaminyi AI, Tikhaze AK, Ageev FT, Kukharchuk V, Belenkov YN. Oxidative stress in atherosclerosis and diabetes. *Bull Exp Biol Med*. 2005; 140:41–3.
55. Birlouez-Aragon I, Saavedra G, Tessier FJ, Galinier A, Ait-Ameur L, Lacoste F, Niamba CN, Alt N, Somoza V, Lecerf JM. A diet based on high-heat-treated foods promotes risk factors for diabetes mellitus and cardiovascular diseases. *Am J Clin Nutr*. 2010;91:1220–6.
56. Drewnowski A, Almiron-Roig E. Human perceptions and preferences for fat-rich foods. In: Montmayeur JP, le Coutre J, editors. *Fat detection: taste, texture, and post ingestive effects*. Boca Raton (FL): CRC Press; 2010. p. 265–89.
57. Lawrence GD, Mavi A, Meral K. Promotion by phosphate of Fe(III)- and Cu(II)-catalyzed autoxidation of fructose. *Carbohydr Res*. 2008; 343:626–35.
58. Berlanga J, Cibrian D, Guillen I, Freyre F, Alba JS, Lopez-Saura P, Merino N, Aldama A, Quintela AM, Triana ME, et al. Methylglyoxal administration induces diabetes-like microvascular changes and perturbs the healing process of cutaneous wounds. *Clin Sci (Lond)*. 2005; 109:83–95.
59. Ogawa S, Nakayama K, Nakayama M, Mori T, Matsushima M, Okamura M, Senda M, Nako K, Miyata T, Ito S. Methylglyoxal is a predictor in type 2 diabetic patients of intima-media thickening and elevation of blood pressure. *Hypertension*. 2010;56:471–6.

60. Yu PH. Deamination of methylamine and angiopathy; toxicity of formaldehyde, oxidative stress and relevance to protein glycooxidation in diabetes. *J Neural Transm Suppl.* 1998;52:201–16.
61. Mozaffarian D, Micha R, Wallace S. Effects on coronary heart disease of increasing polyunsaturated fat in place of saturated fat: a systematic review and meta-analysis of randomized controlled trials. *PLoS Med.* 2010;7:e1000252.
62. Ramsden CE, Hibbeln JR, Majchrzak SF, Davis JM. n-6 fatty acid-specific and mixed polyunsaturated dietary interventions have different effects on CHD risk: a meta-analysis of randomised controlled trials. *Br J Nutr.* 2010;104:1586–600.
63. Strayer D, Belcher M, Dawson T, Delaney B, Fine J, Flickinger B, Friedman P, Heckel C, Hughes J, Kincs F, et al. *Food fats and oils.* Washington, DC: Institute of Shortening and Edible Oils, Inc, 2006:44.
64. Mente A, de Koning L, Shannon HS, Anand SS. A systematic review of the evidence supporting a causal link between dietary factors and coronary heart disease. *Arch Intern Med.* 2009;169:659–69.
65. Yavin E, Brand A, Green P. Docosahexaenoic acid abundance in the brain: a biodevice to combat oxidative stress. *Nutr Neurosci.* 2002;5:149–57.
66. Romieu I, Garcia-Esteban R, Sunyer J, Rios C, Alcaraz-Zubeldia M, Velasco SR, Holguin F. The effect of supplementation with omega-3 polyunsaturated fatty acids on markers of oxidative stress in elderly exposed to PM(2.5). *Environ Health Perspect.* 2008;116:1237–42.
67. Kuczynski B, Reo NV. Evidence that plasmalogen is protective against oxidative stress in the rat brain. *Neurochem Res.* 2006;31:639–56.
68. Lawrence GD. *The fats of life: essential fatty acids in health and disease.* Piscataway (NJ): Rutgers University Press, 2010.
69. Prickett JD, Trentham DE, Robinson DR. Dietary fish oil augments the induction of arthritis in rats immunized with type II collagen. *J Immunol.* 1984;132:725–9.
70. Lawrence GD. Effect of dietary lipids on adjuvant-induced arthritis in rats. *Nutr Res.* 1990;10:283–90.
71. Galarraga B, Ho M, Youssef HM, Hill A, McMahon H, Hall C, Ogston S, Nuki G, Belch JJ. Cod liver oil (n-3 fatty acids) as a non-steroidal anti-inflammatory drug sparing agent in rheumatoid arthritis. *Rheumatology (Oxford).* 2008;47:665–9.
72. Lau CS, Morley KD, Belch JJ. Effects of fish oil supplementation on non-steroidal anti-inflammatory drug requirement in patients with mild rheumatoid arthritis—a double-blind placebo controlled study. *Br J Rheumatol.* 1993;32:982–9.
73. Kremer JM, Lawrence DA, Petrillo GF, Litts LL, Mullaly PM, Rynes RI, Stocker RP, Parhami N, Greenstein NS, Fuchs BR, et al. Effects of high-dose fish oil on rheumatoid arthritis after stopping nonsteroidal antiinflammatory drugs. Clinical and immune correlates. *Arthritis Rheum.* 1995;38:1107–14.
74. Cleland LG, James MJ. Rheumatoid arthritis and the balance of dietary N-6 and N-3 essential fatty acids. *Br J Rheumatol.* 1997;36:513–4.
75. Calder PC. Session 3: Joint Nutrition Society and Irish Nutrition and Dietetic Institute Symposium on 'Nutrition and autoimmune disease' PUFA, inflammatory processes and rheumatoid arthritis. *Proc Nutr Soc.* 2008;67:409–18.
76. Calder PC. n-3 polyunsaturated fatty acids, inflammation, and inflammatory diseases. *Am J Clin Nutr.* 2006;83:1505S–19S.
77. Broughton KS, Johnson CS, Pace BK, Liebman M, Kleppinger KM. Reduced asthma symptoms with n-3 fatty acid ingestion are related to 5-series leukotriene production. *Am J Clin Nutr.* 1997;65:1011–7.
78. Nagakura T, Matsuda S, Shichijyo K, Sugimoto H, Hata K. Dietary supplementation with fish oil rich in omega-3 polyunsaturated fatty acids in children with bronchial asthma. *Eur Respir J.* 2000;16:861–5.
79. Thornley S, Stewart A, Marshall R, Jackson R. Per capita sugar consumption is associated with severe childhood asthma: an ecological study of 53 countries. *Prim Care Respir J.* 2011;20:75–8.
80. Macrae FA. Fat and calories in colon and breast cancer: from animal studies to controlled clinical trials. *Prev Med.* 1993;22:750–66.
81. Weisburger JH. Dietary fat and risk of chronic disease: mechanistic insights from experimental studies. *J Am Diet Assoc.* 1997;97:516–23.
82. Braden LM, Carroll KK. Dietary polyunsaturated fat in relation to mammary carcinogenesis in rats. *Lipids.* 1986;21:285–8.
83. Carroll KK, Braden LM, Bell JA, Kalamegham R. Fat and cancer. *Cancer.* 1986;58:1818–25.
84. Zock PL, Katan MB. Linoleic acid intake and cancer risk: a review and meta-analysis. *Am J Clin Nutr.* 1998;68:142–53.
85. Williams CD, Whitley BM, Hoyo C, Grant DJ, Iraggi JD, Newman KA, Gerber L, Taylor LA, McKeever MG, Freedland SJ. A high ratio of dietary n-6/n-3 polyunsaturated fatty acids is associated with increased risk of prostate cancer. *Nutr Res.* 2011;31:1–8.
86. Wirfält E, Mattisson I, Gullberg B, Olsson H, Berglund G. Fat from different foods show diverging relations with breast cancer risk in postmenopausal women. *Nutr Cancer.* 2005;53:135–43.
87. Bougnoux P, Giraudeau B, Couet C. Diet, cancer, and the lipidome. *Cancer Epidemiol Biomarkers Prev.* 2006;15:416–21.
88. Woutersen RA, Appel MJ, van Garderen-Hoetmer A, Wijnands MV. Dietary fat and carcinogenesis. *Mutat Res.* 1999;443:111–27.
89. Wang D, Dubois RN. Eicosanoids and cancer. *Nat Rev Cancer.* 2010;10:181–93.
90. Harris RE. Cyclooxygenase-2 (cox-2) blockade in the chemoprevention of cancers of the colon, breast, prostate, and lung. *Inflammopharmacology.* 2009;17:55–67.
91. Slaga TJ, Klein-Szanto AJ, Triplett LL, Yotti LP, Trosko KE. Skin tumor-promoting activity of benzoyl peroxide, a widely used free radical-generating compound. *Science.* 1981;213:1023–5.
92. Norat T, Bingham S, Ferrari P, Slimani N, Jenab M, Mazuir M, Overvad K, Olsen A, Tjønneland A, Clavel F, et al. Meat, fish, and colorectal cancer risk: the European Prospective Investigation into cancer and nutrition. *J Natl Cancer Inst.* 2005;97:906–16.
93. Wood JD, Enser M, Fisher AV, Nute GR, Sheard PR, Richardson RI, Hughes SI, Whittington FM. Fat deposition, fatty acid composition and meat quality: a review. *Meat Sci.* 2008;78:343–58.
94. Smith SB, Gill CA, Lunt DK, Brooks MA. Regulation of fat and fatty acid composition in beef cattle. *Asian-Aust J Anim Sci.* 2009;22:1225–33.
95. Lijinsky W. Nitrosamines and nitrosamides in the etiology of gastrointestinal cancer. *Cancer.* 1977;40:2446–9.
96. Alexander DD, Morimoto LM, Mink PJ, Cushing CA. A review and meta-analysis of red and processed meat consumption and breast cancer. *Nutr Res Rev.* 2010;23:349–65.
97. Alexander DD, Mink PJ, Cushing CA, Scourman B. A review and meta-analysis of prospective studies of red and processed meat intake and prostate cancer. *Nutr J.* 2010;9:50.
98. Micha R, Wallace SK, Mozaffarian D. Red and processed meat consumption and risk of incident coronary heart disease, stroke, and diabetes mellitus: a systematic review and meta-analysis. *Circulation.* 2010;121:2271–83.
99. Astrup A, Dyerberg J, Elwood P, Hermansen K, Hu FB, Jakobsen MU, Kok FJ, Krauss RM, Lecerf JM, Legrand P, et al. The role of reducing intakes of saturated fat in the prevention of cardiovascular disease: where does the evidence stand in 2010? *Am J Clin Nutr.* 2011;93:684–8.
100. German JB, Gibson RA, Krauss RM, Nestel P, Lamarche B, van Staveren WA, Steijns JM, de Groot LC, Lock AL, Destaillets F. A reappraisal of the impact of dairy foods and milk fat on cardiovascular disease risk. *Eur J Nutr.* 2009;48:191–203.
101. Huth PJ, Park KM. Influence of dairy product and milk fat consumption on cardiovascular disease risk: a review of the evidence. *Adv Nutr.* 2012;3:266–85.
102. Micha R, Mozaffarian D. Saturated fat and cardiometabolic risk factors, coronary heart disease, stroke, and diabetes: a fresh look at the evidence. *Lipids.* 2010;45:893–905.
103. Volk MG. An examination of the evidence supporting the association of dietary cholesterol and saturated fats with serum cholesterol and development of coronary heart disease. *Altern Med Rev.* 2007;12:228–45.
104. Ng TK, Hayes KC, DeWitt GF, Jegathesan M, Satgunasingam N, Ong AS, Tan D. Dietary palmitic and oleic acids exert similar effects on serum cholesterol and lipoprotein profiles in normocholesterolemic men and women. *J Am Coll Nutr.* 1992;11:383–90.

105. Choudhury N, Tan L, Truswell AS. Comparison of palmolein and olive oil: effects on plasma lipids and vitamin E in young adults. *Am J Clin Nutr.* 1995;61:1043–51.
106. Kaunitz H, Dayrit CS. Coconut oil consumption and coronary heart disease. *Philippine J Coconut Studies.* 1992;17:18–20.
107. German JB, Dillard CJ. Saturated fats: what dietary intake? *Am J Clin Nutr.* 2004;80:550–9.
108. Schuster GS, Dirksen TR, Ciarlone AE, Burnett GW, Reynolds MT, Lankford MT. Anticaries and antiplaque potential of free-fatty acids in vitro and in vivo. *Pharmacol Ther Dent.* 1980;5:25–33.
109. Müller H, Lindman AS, Brantsaeter AL, Pedersen JI. The serum LDL/HDL cholesterol ratio is influenced more favorably by exchanging saturated with unsaturated fat than by reducing saturated fat in the diet of women. *J Nutr.* 2003;133:78–83.
110. Lamarche B. Review of the effect of dairy products on non-lipid risk factors for cardiovascular disease. *J Am Coll Nutr.* 2008;27:741S–6S.
111. Azadbakht L, Mirmiran P, Esmailzadeh A, Azizi F. Dairy consumption is inversely associated with the prevalence of the metabolic syndrome in Tehranian adults. *Am J Clin Nutr.* 2005;82:523–30.
112. van Meijl LE, Vrolix R, Mensink RP. Dairy product consumption and the metabolic syndrome. *Nutr Res Rev.* 2008;21:148–57.
113. Elwood PC, Pickering JE, Givens DI, Gallacher JE. The consumption of milk and dairy foods and the incidence of vascular disease and diabetes: an overview of the evidence. *Lipids.* 2010;45:925–39.
114. Hu FB, Stampfer MJ, Manson JE, Rimm E, Colditz GA, Speizer FE, Hennekens CH, Willett WC. Dietary saturated fats and their food sources in relation to the risk of coronary heart disease in women. *Am J Clin Nutr.* 1999;70:1001–8.
115. Dreon DM, Fernstrom HA, Campos H, Blanche P, Williams PT, Krauss RM. Change in dietary saturated fat intake is correlated with change in mass of large low-density-lipoprotein particles in men. *Am J Clin Nutr.* 1998;67:828–36.
116. Nelson GJ, Schmidt PC, Kelley DS. Low-fat diets do not lower plasma cholesterol levels in healthy men compared to high-fat diets with similar fatty acid composition at constant caloric intake. *Lipids.* 1995;30:969–76.
117. Turley ML, Skeaff CM, Mann JI, Cox B. The effect of a low-fat, high-carbohydrate diet on serum high density lipoprotein cholesterol and triglyceride. *Eur J Clin Nutr.* 1998;52:728–32.
118. Dennison BA, Rockwell HL, Baker SL. Excess fruit juice consumption by preschool-aged children is associated with short stature and obesity. *Pediatrics.* 1997;99:15–22.
119. Dennison BA. Fruit juice consumption by infants and children: a review. *J Am Coll Nutr.* 1996;15:4S–11S.
120. Bray GA, Nielsen SJ, Popkin BM. Consumption of high-fructose corn syrup in beverages may play a role in the epidemic of obesity. *Am J Clin Nutr.* 2004;79:537–43.
121. Elliott SS, Keim NL, Stern JS, Teff K, Havel PJ. Fructose, weight gain, and the insulin resistance syndrome. *Am J Clin Nutr.* 2002;76:911–22.
122. Le Poul E, Loison C, Struyf S, Springael JY, Lannoy V, Decobecq ME, Brezillon S, Dupriez V, Vassart G, Van Damme J, et al. Functional characterization of human receptors for short chain fatty acids and their role in polymorphonuclear cell activation. *J Biol Chem.* 2003;278:25481–9.
123. Liuba P, Persson J, Luoma J, Ylä-Herttuala S, Pesonen E. Acute infections in children are accompanied by oxidative modification of LDL and decrease of HDL cholesterol, and are followed by thickening of carotid intima-media. *Eur Heart J.* 2003;24:515–21.
124. Reiser S, Hallfrisch J, Michaelis OE 4th, Lazar FL, Martin RE, Prather ES. Isocaloric exchange of dietary starch and sucrose in humans. I. Effects on levels of fasting blood lipids. *Am J Clin Nutr.* 1979;32:1659–69.
125. Bantle JP, Raatz SK, Thomas W, Georgopoulos A. Effects of dietary fructose on plasma lipids in healthy subjects. *Am J Clin Nutr.* 2000;72:1128–34.
126. Solyst JT, Michaelis OE 4th, Reiser S, Ellwood KC, Prather ES. Effect of dietary sucrose in humans on blood uric acid, phosphorus, fructose, and lactic acid responses to a sucrose load. *Nutr Metab.* 1980;24:182–8.
127. Sakai M, Oimomi M, Kasuga M. Experimental studies on the role of fructose in the development of diabetic complications. *Kobe J Med Sci.* 2002;48:125–36.
128. Rebolledo A, Rebolledo OR, Marra CA, Garcia ME, Roldan Palomo AR, Rimorini L, Gagliardino JJ. Early alterations in vascular contractility associated to changes in fatty acid composition and oxidative stress markers in perivascular adipose tissue. *Cardiovasc Diabetol.* 2010;9:65–73.
129. Malik VS, Popkin BM, Bray GA, Despres JP, Hu FB. Sugar-sweetened beverages, obesity, type 2 diabetes mellitus, and cardiovascular disease risk. *Circulation.* 2010;121:1356–64.
130. Malik VS, Popkin BM, Bray GA, Despres JP, Willett WC, Hu FB. Sugar-sweetened beverages and risk of metabolic syndrome and type 2 diabetes: a meta-analysis. *Diabetes Care.* 2010;33:2477–83.
131. Lim JS, Mietus-Snyder M, Valente A, Schwarz JM, Lustig RH. The role of fructose in the pathogenesis of NAFLD and the metabolic syndrome. *Nat Rev Gastroenterol Hepatol.* 2010;7:251–64.
132. Cirillo P, Sautin YY, Kanellis J, Kang DH, Gesualdo L, Nakagawa T, Johnson RJ. Systemic inflammation, metabolic syndrome and progressive renal disease. *Nephrol Dial Transplant.* 2009;24:1384–7.
133. Stanhope KL, Schwarz JM, Keim NL, Griffen SC, Bremer AA, Graham JL, Hatcher B, Cox CL, Dyachenko A, Zhang W, et al. Consuming fructose-sweetened, not glucose-sweetened, beverages increases visceral adiposity and lipids and decreases insulin sensitivity in overweight/obese humans. *J Clin Invest.* 2009;119:1322–34.