Review

Fruit and vegetable peels: Paving the way towards the development of new generation therapeutics

Hamendra S. Parmar^{1,*}, Yamini Dixit², Anand Kar²

¹ School of Biotechnology, Devi Ahilya University, Takshashila Campus, Indore, India;

² Thyroid Research Unit, School of Life Sciences, Devi Ahilya University, Takshashila Campus, Indore, India.

ABSTRACT: Cardiovascular diseases (CVDs), diabetes mellitus (DM), cancer, and thyroid abnormalities are major health problems prevalent around the world and are responsible for a large portion of morbidity and mortality out of health problems overall. Advances in genomics and proteomics in recent years have led to an explosion in the number of possible therapeutic targets and drug candidates through use of molecular approaches, chemical synthesis, traditional medicinal chemistry, and phyto-chemistry and through the exploration of novel herbal preparations. However, virtually none of these candidates are devoid of potential adverse drug reaction(s) or undesirable side effects. Therefore, the clear need is to look to alternative ways to develop novel drug candidates with fewer side effects and less cost. Interestingly, the last few years have seen an increase in the number of available reports on fruits and vegetable peels, and particularly on their biological activity, their content of different bioactive compounds, their chemical characterization, understanding of their structureactivity relationships, isolation and purification of commercially important chemicals without using high throughput techniques, etc. Therefore, research in the field of fruit and vegetable peels should present immense possibilities for drug discovery and development of cost-effective therapies that have fewer or practically no side effects. This virtual explosion of interest in fruit and vegetable peels as a source of medicinal and nutritional value has led to the present review.

Keywords: Cardiovascular problems, cancer, diabetes mellitus, thyroid problems, peels

*Address correspondence to:

1. Introduction

Cardiovascular diseases including coronary heart disease (heart attacks), cerebrovascular disease, raised blood pressure (hypertension), peripheral artery disease, rheumatic heart disease, congenital heart disease and heart failure, diabetes mellitus (both types 1 and 2), thyroid abnormalities (broadly hypo- and hyperthyroidism), and cancer are among the most prevalent diseases around the world and are responsible for a large portion of morbidity and mortality out of health problems overall (*1-9*).

A vast body of literature is available on the possible therapeutic targets for those diseases, although many substances are either in clinical trials or used in practice (10-37). However, they have some major drawbacks including their high cost and adverse effects like cardiovascular events, cancer, aging, cardiac and renal toxicity, and increased oxidative stress (38-51). Oxidative stress is itself known to be a root cause for the progression and development of the diseases mentioned (6), and modern medicines should be designed in a way to maintain a healthy homeostasis between oxidants and antioxidants. As most herbal preparations are antioxidative in nature, they improve health directly by reducing oxidative stress (6, 52, 53). In fact, many herbal extracts are known to be antiperoxidative, anti-cancer, cardio-protective, anti-diabetic, and thyro-regulatory in nature (3, 4, 54-58). However, the identification of the plant(s) and its availability, precise chemical composition, dose, potential use for ailment(s), precise mechanism of action, unpredictable toxicity, and cost are major concerns that hinder the use of herbal preparations (59). That said, fruit and vegetable peels have advantages over other herbal extracts, as they are easily identifiable, commonly used by people, rich in various bioactive compounds, and some of their compounds have been characterized in terms of their chemical structures and biological properties through use of structure-activity relationships (SAR). Additionally, peels are usually considered waste, so they are obviously cost-effective (60-64).

Therefore, the present review has attempted to

Dr. Hamendra Singh Parmar, School of Biotechnology, Devi Ahilya University, Takshashila Campus, Khandwa Road, Indore 452001, M.P., India. e-mail: hamendrasingh999@yahoo.co.in

assess the emerging potential of fruit and vegetable peels for use in developing new generation therapeutics.

2. Anti-peroxidative or radical-scavenging properties

Free radical production in any organism can either be accidental or deliberate. Free radicals have increasingly been accepted as commonplace and important biochemical intermediates, leading to these compounds being implicated in a large number of human diseases including cardiovascular problems, diabetes mellitus, cancer, thyroid disorders, and Alzheimer's disease (6).

Various fruit and vegetable peel extracts or compounds are known to be antiperoxidative in nature and their different in vitro or in vivo mechanism(s) have also been reported (Table 1). The antiperoxidative or radical-scavenging potential of the peel extracts from C. sinensis, P. granatum, M. paradisiaca, C. vulgaris, C. melo, and M. indica is well documented in both in vivo and in vitro models (52,53,63-67). In fact, the current authors have demonstrated that these peel extracts work mainly through the direct radical scavenging of various types of radicals in a dose-specific manner (64). Possible mechanism(s) of their antiperoxidative potential might be mediated via the presence of a variety of polyphenols and flavonoids in different concentrations. Specifically, C. sinensis was found to be an efficient scavenger for DPPH, singlet oxygen, and various peroxyradicals. P. granatum and M. paradisiaca were also found to be effective against all the aforementioned radicals and nitric oxide (NO) radicals as well, while *M. indica* was found to be effective only against peroxyradicals and C. vulgaris and C. melo were similarly found to be effective only against singlet oxygen and peroxyradicals. All of the aforementioned peels also have a minor influence on enzymatic and non-enzymatic oxidative defense, which includes catalase (CAT), superoxide dismutase (SOD), and reduced glutathione (GSH), particularly

Table 1. Antiperoxidative potential of fruit and vegetable peels

Botanical name	English name	References
1. Citrus sinensis	Sweet orange	(64-66)
2. Citrus reticulata	Mandarin	(68,149)
3. Citrus paradisi	Jaffa grapefruit	(68)
4. Musa paradisiaca	Banana	(52,64,65,94)
5. Citrullus vulgaris	Watermelon	(63,64,67)
6. Cucumis melo	Melon	(63,64,67)
7. Mangifera indica	Mango	(63,64,67,150,151)
8. Punica granatum	Pomegranate	(52,64,65,99,135)
9. Passiflora liguralis	Sweet granadilla	(153)
10. Legenaria siceraria	Bottle gourd	(62)
11. Solanum melongena L.	Brinjal	(73,74)
12. Solanum tuberosam	Potato	(75,76)
13. Capsicum annuum L.	Sweet pepper	(74)
14. Cydonia vulgaris	Quince	(154)
15. Pyrus malus	Apple	(60,71,133)
16. Pyrus pashia	Pear	(60,71)
17. Prunus persica	Peaches	(71)

in the event of disease (53,66). Other citrus fruit peels including C. reticulata and C. paradisi are also known to have an antiperoxidative effect (68,69). Similarly, extracts of P. liguralis peels are reported to have considerable antioxidant activity, as represented by the trolox equivalent antioxidant capacity (TEAC) value, due to the presence of various antioxidative bioactive compounds, certainly indicating the importance of this peel as an alternative source of bioactive compounds (70). Peach, pear, and apple peels are also reported to have antioxidative potential according to various in vitro methods such as total radical-trapping antioxidative potential (TRAP) values, which also correlate with their polyphenolic content. Peels of those fruits have also been found to be antiperoxidative in hypercholesterolemic diet-fed animals (71). Pears and apples were further characterized by beta-carotene bleaching and NO and DPPH radical-scavenging potential (60). Peels from Red grape marc have also been found to be a radical quencher according to a beta-carotene bleaching assay (72). Solanum melongena is known to contain very strong antioxidants, including nasunin, and its antioxidative potential has been demonstrated using electron spin resonance spectrometric analysis, 5,5-dimethyl-1pyrroline-N-oxide (DMPO), spin trapping, hydroxyl (•OH) or superoxide anion radicals (O²⁻) generated by a Fenton reaction, and hypoxanthine-xanthine oxidase systems (73). Both S. melongena L. and C. annuum L. peels are also reported to have in vitro antiperoxidative potential due to the presence of some other strong antioxidant compounds (74). Solanum tuberosum peel extract has also been found to have an antioxidative effect on erythrocytes and in rats with streptozotocininduced diabetes (75,76). Jaffa grapefruit peels have been evaluated in DPPH and beta-carotene linoleate model systems and have been found to be radical scavengers in vitro (77). The peel extract of L. siceraria has also recently been reported to have antiperoxidative potential in both in vitro and in vivo studies; in vitro analysis demonstrated that this peel extract not only quenches DPPH radicals but also lowers hepatic lipid peroxidation values induced by CCl_4 and H_2O_2 (62). A parallel *in vivo* study on normal healthy and hyperthyroid mice further confirmed its antioxidative efficacy (62).

3. Cardiovascular protective effect

Many flavonoids and their glycosides present in herbal extracts are known for their cardiovascular regulatory properties (Table 2) and many are abundantly available in fruit and/or vegetable peels including rutin, isoquercetin, narirutin, narcissin, quercetin, kaempferol, luteolin, and apigenin are known to have a vasodilatory and hypotensive effect (78-80). Some of the flavonoids, such as quercetin and quercetin glycosides, are reported to have lipid-lowering and anti-atherosclerotic activity (79,81-83). In fact, hesperidin and naringin, both citrus

 Table 2. Fruit and vegetable peels known for their cardiovascular effect

Botanical name	English name	References
1. Citrus sinensis	Sweet orange	(52)
2. Citrus reticulata	Mandarin	(84)
3. Musa paradisiaca	Banana	(52)
4. Citrullus vulgaris	Watermelon	(65,118)
5. Cucumis melo	Melon	(65)
6. Mangifera indica	Mango	(52,115)
7. Punica granatum	Pomegranate	(52)
8. Citrus paradisi	Jaffa grapefruit	(77)
9. Pyrus malus	Apple	(60,71)
10. Pyrus pashia	Pear	(60,71)
11. Prunus persica	Peaches	(60)

bioflavonoids also present in citrus fruit peels, exhibit biological and pharmacological properties, such as anti-inflammatory, lipid-lowering, and antioxidative behavior; all are related to cardiovascular health (84,85).

The mechanism(s) of the aforementioned effects may be explained by the fact that oxidative modification of low-density lipoproteins (LDL) by free radicals is an early event in the pathogenesis of atherosclerosis. The rapid uptake of oxidatively modified LDL via a scavenger receptor leads to the formation of foam cells. Oxidized LDL also has a number of other atherogenic properties. A number of mechanisms are likely to contribute to inhibition of LDL oxidation by flavonoids. Flavonoids may directly scavenge some radical species by acting as chain-breaking antioxidants (86). In addition, they may recycle other chain-breaking antioxidants such as α -tocopherol by donating a hydrogen atom to the tocopheryl radical (87). Transition metals such as iron and copper are important pro-oxidants, and some flavonoids can chelate divalent metal ions, hence preventing free radical formation.

A detailed in vivo study of C. sinensis, P. granatum, M. paradisiaca, C. vulgaris, C. melo, and M. indica peels in a diet-induced animal model of atherosclerosis revealed the anti-atherogenic potential of extracts. The study also revealed their direct benefit of maintaining cardiovascular health by positively influencing serum lipids (including total cholesterol, triglycerides, LDLcholesterol, and VLDL-cholesterol), the atherogenic index, glucose, tissue lipid peroxidation, the serum level of creatinine kinase-MB enzyme, and histopathological alterations (52,67). The possible reasons for this beneficial role correlated with the presence of a variety of total flavonoids, phenolic compounds, and ascorbic acid content of the peel extracts (66,67). The aforementioned fruit peels are specifically known to contain various bioactive compounds that are already known for their cardiovascular or related benefits, including antiperoxidative, anti-inflammatory, and cardioprotective action. In brief, the protective effect of C. sinensis peels might be due to the presence of polymethoxylated flavones, C-glycosylated flavones, O-glycosylated flavones, flavonols, phenolic acids, nobiletin, hesperidin,

 Table 3. Anti-diabetic or gluco-regulatory potential of fruit and vegetable peels

Botanical name	English name	References
1. Citrus sinensis	Sweet orange	(65-67,119)
2. Punica granatum	Pomegranate	(65,67)
3. Mangifera indica	Mango	(63)
4. Citrullus vulgaris	Watermelon	(63)
5. Solanum tuberosum	Potato	(75)
6. Legenaria siceraria	Bottle gourd	(62)

and naringin (88-92). In M. paradisiaca, dopamine seems to be responsible as it is known to have strong antiperoxidative properties that are known to be associated with the amelioration of cardiovascular problem(s) (6,93-97). In P. granatum, some compounds are already known for their antiperoxidative and antiinflammatory properties, including oleanolic, ursolic, and gallic acids, punicalagin, ellagitannin, ellagic acid, and catechin (98-107). Similarly, the anti-atherogenic activity of the peel extract of M. indica could be the result of the action of its rich polyphenolic content. Q 3-galactoside, Q 3-glucoside, and Q 3-arabinoside, gallic acid, and mangiferin are reported to have an antioxidative, antiinflammatory, and cardioprotective role (108-115). The protective activity of C. vulgaris and C. melo peels mainly relates to their high content of citrulline, an essential amino acid that helps in nitric oxide synthesis that, in turn, enhances vasodilatation (116-118). Peels from Jaffa grapefruit (C. paradisi), pears, peaches, and apples were also evaluated for their possible cardiovascular benefits in hypercholesterolemic dietfed animals. These peels increased plasma antioxidant capacity and improved plasma levels of different lipids. Further correlation studies revealed that the observed benefits of these peels might be mediated via the presence of total flavonoids, phenolics, phenolic acids, and dietary fiber at various levels of correlation (77).

4. Antidiabetic or gluco-regulatory potential of fruit and vegetable peels

Dietary antioxidant compounds such as bio-flavonoids may offer some protection against the early stage of diabetes mellitus and the development of complications (Table 3). Available reports describe the known mechanism(s) of bioflavonoids that are present in peels, such as hesperidin and naringin as are present in citrus fruit peels. These peels play an antidiabetic role in C57BL/KsJ-db/db mice *via* regulation of glucoregulatory enzymes *i.e.*, they decrease the activity of glucose-6-phosphatase and phosphoenol pyruvate with a concomitant increase in the activity of hepatic glucokinase, increased hepatic glycogen content, and increased serum insulin along with a decrease in serum glucose concentrations (80).

C. sinensis and P. granatum peel extracts have also been found to be thyroid-stimulating in nature

when evaluated in normal healthy animals (65). Their antidiabetic potential was further confirmed by the experimentation using alloxan induced diabetes mellitus, hypercholesterolemic diet fed, and hyperthyroid animal models of study, which indentified the mechanism for the observed effects of C. sinensis peels and suggested that the antidiabetic potential of this peel extract might be mediated via antiperoxidation, α -amylase enzyme activity inhibition that is responsible for the conversion of complex carbohydrates to glucose, increased hepatic glycogen content, insulin-stimulating activity, and repair of secretory defects in β -cells (53). Inhibition of α -amylase enzyme activity by Citrus sinensis peel extract was also reported by other authors (119). P. granatum is suggested to have intrinsic antiperoxidative and hypoglycemic properties that may be attributed to some of its bioactive compounds, including oleanolic, ursolic, and gallic acids, punicalagin, ellagitannin, ellagic acid, and catechin (98-107). This protective effect was further correlated with the total phenolic and flavonoid compound content in the peels (53). However, M. indica and C. vulgaris have displayed neither any intrinsic hypoglycemic potential nor any antidiabetic potential in diabetic models (data not shown) but have been found to produce hypoglycemia in hyperlipidemiainduced diabetes (67). Therefore, these peel extracts may work via gluconeogenesis or glycogenolysis or glucose uptake in hypercholesterolemic animals. Therefore, it seems that these peels may be beneficial in obesity induced type 2 diabetic condition or in metabolic syndrome. Antidiabetic role of potato or S. tuberosum peels against streptozotocin induced diabetic model was also reported where, reversal in almost all the diabetic changes including serum glucose, body weight, polydipsia, polyuria, elevated activity of serum transaminases (ALT and AST) and hepatic MDA levels, and reduced glutathione (GSH) was observed (75). However, the plausible mechanism for this antidiabetic effect has not been completely elucidated but antiperoxidative potential was presumably a major contributing factor to the effect observed. Similarly, the peel extract of L. siceraria has been found to cause hypoglycemia in normal healthy and hyperthyroid mice. The hypoglycemic potential observed might be the outcome of thyroid and glucose-6-phosphatase inhibitory activity of the peel extract (62).

5. Thyro-regulatory potential

Some plant compounds are already known to influence the thyroid hormone homeostasis at various levels, including that of binding of TSH-receptor, thyroid-iodide transport and conversion of T_4 to T_3 (120). However, few reports (Table 4) have demonstrated the thyro-regulatory potential of fruit and vegetable peels (52,53,62-67). Peels from C. sinensis and M. paradisiaca have been found to inhibit the thyroid. Where, reduction in both the thyroid hormones was observed, in response to either of the peel extract. Therefore, it was suggested that both C. sinensis and M. paradisiaca might be inhibiting thyroid hormones not only at glandular level, but also at the level of peripheral conversion of T_4 to T_3 . The antithyroidal role of C. sinensis might be mediated through the inhibition of thyroid peroxidase (TPO); the key enzyme in thyroid hormone biosynthesis, as it contains the phenolic compound naringin which inhibits the activity of TPO (121-123). Similarly, the antithyroidal role of M. paradisiaca might be mediated by its high dopamine content, which is already known to inhibit the thyroid as previously indicated (94,124,125).

The peel extracts of *M. indica*, *C. vulgaris*, and *C. Melo* were found to be thyro-stimulatory in nature. This thyroid stimulatory nature was further confirmed by a study of rats with chemically-induced hypothyroidism in which the administration of test peel extracts restored the serum levels of two thyroid hormones to normal in hypothyroid animals (*63*). These results clearly demonstrated the role the aforementioned peel extracts had in ameliorating hypothyroidism. An increased level of both thyroid hormones T_3 and T_4 demonstrated the thyroid stimulatory potential of these peel extracts on both the glandular level (the only source for T_4 synthesis) and at the level of T_3 .

Thus far, the mechanism for the effect(s) observed may relate to the presence of various small polyphenolic molecules that might play a major role in thyroid stimulatory activity, as they are already reported to influence thyroid hormone metabolism at genomic level. For instance, they increase the activity of the type 2 iodothyronine deiodinase gene (126). Secondly, other mechanisms are also possible, such as TPO stimulation, enhanced glandular functionality, and 5'-deiodinase activity, and could not be ruled out by these studies.

Botanical name	English name	Nature	References
1. Citrus sinensis	Sweet orange	Thyroid-inhibiting	(65,66)
2. Musa paradisiaca	Banana	Thyroid-inhibiting	(65)
3. Legenaria siceraria	Bottle gourd	Thyroid-inhibiting	(62)
4. Citrullus vulgaris	Watermelon	Thyroid-stimulating	(63)
5. Cucumis melo	Melon	Thyroid-stimulating	(64)
6. Mangifera indica	Mango	Thyroid-stimulating	(63)

6. Anticancer potential

Cancer is one of the most devastating diseases for which various remedies have been reported, but development of suitable therapeutics to treat this disease is still a major challenge for biomedical professionals. In the search for novel therapies and exploration of hitherto unknown compounds with anticancer potential, some reports have described herbal preparations including fruit and vegetable peels (Table 5) (127-132). A few important biochemical in vitro studies using different cancer cell lines and *in vivo* studies have revealed the potential some fruit peels have to combat variety of cancers, including cancer of the liver, colon, breast, and lung. Peels from different varieties of apples (Rome Beauty, Idared, Cortland, and Golden Delicious) are reported to have an antiproliferative effect (130). Golden delicious apple peels have been reported to inhibit the cell proliferation of HepG2 human liver cancer cells and MCF-7 human breast cancer cells (133). Some of the active principles present in these peels, such as quercetin and quercetin-3-O-beta-D-glucopyranoside, have been found to be responsible for the anticancer activity observed (130,134). The anticancer effect(s) observed might be mediated via inhibition of NF-kappa B activation (131). Some of the triterpenoids are also present in apple peels, including 2 alpha-hydroxyursolic acid, 2 alpha-hydroxy-3 beta-{[(2E)-3-phenyl-1-oxo-2-propenyl]oxy}olean-12-en-28-oic acid, 3 beta-trans-p-coumaroyloxy-2 alpha-hydroxyolean-12-en-28-oic acid, and 2 alphahydroxyursolic acid, and are known to possess anticancer potential via the inhibition of NF-kappa B activation (134). Similarly, peels of Punica granatum are also thought to have an anticancer effect in inflammationassociated cancers (135). Different Citrus varieties including C. reticulata, C. unshiu, and C. natsudaidai are known to prevent tumorigenesis (136,137). C. reticulata peels have displayed potent tumor-suppressing activity in SNU-C4 human colon cancer cells; the mechanism for this is believed to be via the up-regulation of the proapoptotic gene Bax and apoptotic gene caspase-3 along with a concomitant decrease in the expression of the antiapoptotic gene bcl-2 (138).

The peel extract of *C. natsudaidai* has also been demonstrated to act on tumors in B-16 mouse

melanoma and human lung carcinoma cells; it is theorized to contain hydrophobic antitumor compounds (137). In fact, 78 species of the genus *Citrus* are known to inhibit the Epstein-Barr virus early antigen (EBV-EA) activation (responsible for some cancers, including Burkitt's lymphoma) induced by 12-O-tetradecanoylphorbol 13-acetate (TPA); this serves as a useful screening method for anti-tumor promoters and further underscores the importance of peels in the development of potential anti-tumor therapies (139).

Isolated fractions of *D. kaki* (persimmon) peels have potent cytotoxic activity against human oral squamous cell carcinoma cells (HSC-2) and human submandibular gland tumor (HSG) cells (*140*). Interestingly, these fractions also had activity to reverse multiple drug resistance (MDR), further encouraging research on persimmon peels in the prevention and treatment of cancers as MDR is a frequently occurring event in most of the available cancer therapies (*140*). Similarly, cytotoxic and MDR reversal activity were also reported in response to treatment with Feijoa peel extract (*141*).

7. Active principles available in peels and their potential health benefits

Various compounds are present in both vegetable and fruit peels and are known for their different biological activities; these compounds are thought to be the active principles in these peels (Table 6). Different species of S. melongena contain various anthocyanins such as delphinidin 3-(p-coumaroylrutinoside)-5-glucoside (nasunin), delphinidin 3-rutinoside, delphinidin 3-glucoside, and petunidin 3-(p-coumaroylrutinoside)-5-glucoside (petunidin 3RGc5G). These compounds are all reported to have a varying degree of radicalscavenging potential. Delphinidin 3RGcaf5G is reported to have the highest level of radical-scavenging activity in 1,1-diphenyl-2-picrylhydrazyl (DPPH) radical and linoleic acid radical systems, followed by nasunin and petunidin 3RGc5G, in that order (73,142). Interestingly, an ex vivo angiogenesis assay using a rat aortic ring revealed the antiangiogenic and antioxidative potential of nasunin (78). Similarly, delphinidin-3-rutinoside from S. melongena and delphinidin-3-trans-coumaroylrutinoside-5-glucoside from C. annuum L. are also reported to

Table 5. Fruit and vegetable peels known for their anticancer efficacy	Table 5. Fruit and	vegetable p	eels known fo	r their anticancer	efficacy
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Botanical name	English name	References
1. Pyrus malus	Apple	(130,131,133,134)
2. Punica granatum	Pomegranat	(135)
3. Solanum lycopersicum	Tomato	(155)
4. Citrus reticulata blanco	Mandarin orange	(138)
5. Citrus unshiu	Mikan	(136,137)
6. Citri Reticulatae Viride	Green Tangerine Orange	(136,137)
7. Citrus natsudaidai	Japanese summer grape fruit	(137)
8. Diospyros kaki	Persimmon	(140)
9. Feijoa sellowiana	Feijoa	(141)

Name	Source	Biological activity	References
1. Different varieties of delphinidin anthocyanins, delphinidin-3-rutinoside, nasunin	Solanum melongena	Antioxidant	(73,74,78,142)
 Flavonoids including quercetin-3-O-beta-D- glucopyranoside, quercetin-3-O-beta-D-galactopyranoside, quercetin, (-)-catechin, (-)-epicatechin, quercetin-3-O- alpha-L-arabinofuranoside, 2 alpha-hydroxyursolic acid 	Pyrus malus	Anticancer	(61,130,134)
 Various triterpenoids including ursolic acid, 3 beta-<i>trans-</i> p-coumaroyloxy-2 alpha-hydroxyolean-12-en-28-oic acid, (-)-epicatechin, procyanidin B2, chlorogenic acid, and catechins and flavonol glycosides, especially rutin 	Pyrus malus	Antioxidant	(61,130,134)
4. Epicatechin, gallic, and <i>p</i> -coumaric acids	Diospyros kaki	Antiatherosclerotic	(143)
5. Caffeic, <i>p</i> -coumaric, and ferulic acids	Pyrus malus Pyrus malus Pyrus pashia Prunus persica	Lipid lowering	(144,148)
6. Mangiferin, penta- <i>O</i> -galloyl-glucoside, gallic acid, methyl gallate, quercetin <i>O</i> -glycosides, kaempferol <i>O</i> -glycoside, xanthone <i>C</i> -glycosides, mangiferin, isomangiferin, gallotannins	Mangifera indica L.	Antioxidant	(150,151)
 Resorcinols including 5-(11'Z-Heptadecenyl)-resorcinol, 5-(8'Z, 11'Z-Heptadecadienyl)-resorcinol 	Mangifera indica L.	Anti-inflammatory	(152)
8. Fatty acid esters of hydroxybenzoic acid, fatty acid esters of hydroxybenzaldehyde, glucosides of aromatic acids, chlorogenic acids, flavonols, and benzylamine	Cydonia vulgaris	Antioxidant	(154)
9. Xyloglucan (carbohydrate)	Passiflora liguralis	Antioxidant	
10. Flavanon glycosides hesperidin and naringin aglycones hesperetin and naringenin	Some Citrus fruits	Antioxidant	(144,148)
5-Hydroxy-3,6,7,8,3',4'-hexamethoxyflavone	Citrus sinensis	Anticancer	(136,145)
11. Cyclonatsudamine A	Citrus natsudaidai	Vasodilatation	(146)
12. Naringin, naringenin, hesperidin, hesperetin, rutin, nobiletin, and tangeretin	Some Citrus fruit peels	NO radical inhibition	(149)
13. Delphinidin-3-trans-coumaroylrutinoside-5-glucoside	Capsicum annuum L.	Antioxidant	(74)
14. Lycopene and carotenoids	Solanum lycopersicum	Cancer prevention	(155)
15. Hesperidin	Citrus unshiu	Decreased plasma triglycerides	(148)
16. Auraptene and umbelliferone	Citrus natsudaidai	Anticancer	(136)

Table 6. Active principles isolated from fruit and/or vegetable peels and known for their various biological properties

be antiperoxidative according to two different *in vitro* antioxidant capacity assessment assays (74). Similarly, apple peels contain a number of major flavonoids, including quercetin-3-O-beta-D-glucopyranoside, quercetin-3-O-beta-D-galactopyranoside, and trace amounts of quercetin, (–)-catechin, (–)-epicatechin, and quercetin-3-O-alpha-L-arabinofuranoside (130). Among the compounds isolated, quercetin and quercetin-3-O-beta-D-glucopyranoside had potent antioxidative and antiproliferative activity against HepG2 (liver) and MCF-7 (breast) cancer cells, while caffeic acid, quercetin, and quercetin-3-O-beta-D-arabinofuranoside, all phenolic compounds, also had antioxidant activity

(134). Interestingly, most of the flavonoids and phenolic compounds tested were found to be stronger antioxidants when compared to ascorbic acid and might be directly responsible for the antioxidative and antiproliferative activity of apple peels. The presence of triterpenoids, including 2 alpha-hydroxyursolic acid and ursolic acid, in apple peels also makes them a potent cytotoxic or anticancer agent, as evidenced by their inhibitory activity against four tumor cell lines (HL-60, BGC, Bel-7402, and Hela) (61). These compounds have anticancer activity *via* inhibition of NF-kappa B activation. Interestingly, structure-activity relationships (SAR) revealed that these triterpenes possess two hydrogen bond-forming

groups (an H-donor and a carbonyl group) at positions 3 and 28 with cytotoxic activity. The configuration at C-3 was found to be important for anticancer activity, as introduction of an amino group was found to greatly increase cytotoxicity. Other evidence has also confirmed the importance of the C-3 and 28 positions, *e.g.* a 3 beta-amino derivative had 20 times the potency of its parent ursolic acid and 28-aminoalkyl dimer compounds had selective cytotoxicity (*61*).

The peels of persimmons and apples have also been recommended as part of an antiatherosclerotic diet due to their rich amounts of total, soluble, and insoluble dietary fiber, total phenols, epicatechin, and gallic and *p*-coumaric acids along with concentrations of Na, K, Mg, Ca, Fe, and Mn (*143*).

The different active components isolated from citrus fruit peels are also known for their various health benefits and disease protection, including antiperoxidative and anticancer activity, vasodilatation, decreased serum triglycerides, and improved cardiovascular health (144-148). Caffeic acid, p-coumaric acid, ferulic acid, and *p*-hydroxybenzoic acid isolated from *C. unshiu* Marc. peels had antioxidative or radical-scavenging properties as represented by trolox equivalent antioxidant capacity (TEAC) values. Similarly, hesperidin isolated from the same citrus variety was found to decrease the level of serum triglycerides (144,148). A polymethoxyflavone compound, 5-hydroxy-3,6,7,8,3',4'-hexamethoxyflavone (5-OH-HxMF), from the sweet orange (C. sinensis) is found exclusively in the Citrus genus and known for its anticancer and anti-inflammatory potential according to 12-O-tetradecanoylphorbol-13-acetate (TPA)-induced expression of inducible nitric oxide synthase (iNOS) and cyclooxygenase-2 (COX-2), which lead to tumor progression, in mouse skin (145). Pre-treatment with a topical application of 5-OH-HxMF has been found to inhibit the TPA-induced nuclear translocation of nuclear factor-kappa B (NF-kappa B) subunit and DNA binding by blocking phosphorylation of inhibitor kappa B (IkappaB) alpha and p65 and subsequent degradation of IkappaB alpha (145). Another study also suggested the potential of OH-HxMF to inhibit 7,12-dimethylbenz[a] anthracene/TPA-induced skin tumor formation, as evidenced by a reduction in tumor incidence and tumor multiplicity of papillomas at 20 weeks (136). Because of its anti-inflammatory and anti-tumor properties, 5-OH-HxMF may prove to be a novel functional agent to prevent inflammation-associated tumorigenesis (145). A novel compound, cyclonatsudamine A, was isolated from C. natsudaidai and evaluated for its vasodilatory potential in a rat aorta model with norepinephrineinduced contractions. The mechanism of vasodilatation was presumably mediated by increased NO release from endothelial cells (146).

In fact, some citrus fruit peel extracts have been reported to have varying degrees of NO radicalscavenging activity, and these levels have been further correlated with the content of some flavonoids, including naringin, naringenin, hesperidin, hesperetin, rutin, nobiletin, and tangeretin (149). M. indica peels contain compounds such as quercetin O-glycosides, kaempferol O-glycoside, xanthone C-glycosides, mangiferin, and isomangiferin that may serve as natural antioxidants or functional food ingredients (150). Other components, including mangiferin, penta-O-galloyl-glucoside, gallic acid, and methyl gallate, are already reported to scavenge DPPH radicals, suggesting radical-scavenging activity (151). Similarly, two other compounds, 5-(11'Z-heptadecenyl)-resorcinol and 5-(8'Z,11'Zheptadecadienyl)-resorcinol, are also reported to exhibit potent cyclooxygenase-1 (COX-1) and COX-2 inhibitory activity (152). Understanding structure-activity relationships revealed that the degree of unsaturation in the alkyl chain plays a key role in this COX inhibitory activity (152). In a TEAC assay system, an unknown polysaccharide xyloglucan from Passiflora liguralis or granadilla fruit peels was also reported to have antioxidative potential (153). Reports on assessing the capacity to scavenge the 2,2'-diphenyl-1-picrylhydrazyl (DPPH) radical and anion superoxide radical and to induce the reduction of Mo(VI) to Mo(V) indicated that various chlorogenic acids and the flavonols isolated from the peels of Cydonia vulgaris have antioxidative and radical-scavenging properties greater than those of alpha-tocopherol and ascorbic acid (154). Tomato consumption is associated with a lower incidence of upper aerodigestive tract and prostate cancers due to presence of carotenoids, lycopene, and/or beta-carotene, but interestingly the content of these compounds is higher in peels than in other parts of the fruit (155). In fact, an in vitro digestion model using human intestinal cells (Caco-2) revealed that tomato paste enriched with 6% peel increased lycopene absorption into intestinal cells 75% and beta-carotene absorption 41%, clearly demonstrating the important role that active principles present in peels might play in cancer prevention (155).

8. Future scenario

Reviewing all of the findings on fruit and vegetable peels leads to the conclusion that fruit and vegetable peels have immense potential as novel and promising therapies against the most prevalent diseases, *i.e.*, cardiovascular problems, diabetes mellitus, thyroid abnormalities, and various cancers.

Interestingly, most peels are considered to be waste and are believed to adversely affect the cleanliness of urban areas, so their utilization in pharma or nutraceuticals will certainly offer the potential for costeffective new generation therapeutics and also enhance the value of fruits and vegetables. As oxidative stress is one of the major factors responsible for various diseases and tissue damage, the presence of strong antioxidants in peels suggests a reduced likelihood of potential drug toxicity or adverse drug reaction(s). However, well planned pre-clinical studies exploring toxicity and efficacy evaluations that provide an understanding of molecular pathways of the biological effects observed are still needed before any clinical trials can be conducted.

Based on the available literature, evidence, and first-hand experience working in this field, the current authors are quite optimistic that the path from fruit and vegetable peels used in the laboratory to peels available on the market will only take a few more years; soon, they may serve as new generation therapeutics to treat cancer, cardiovascular diseases, diabetes mellitus and thyroid abnormalities.

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References

- Smletzer SG, Bare BG. Brunner and Suddhath's Text Book of Medical-Surgical Nursing. 7th ed., J.B. Lippincott, Philadelphia, PA, USA, 1992.
- Chockalingam A, Chalmers J, Lisheng L, Labarthe D, MacMahon S, Martin I, Whitworth J. Prevention of cardiovascular diseases in developing countries: Agenda for action (statement from a WHO-ISH Meeting in Beijing, October 1999). J Hypertens. 2000; 18:1705-1708.
- Kar A, Panda S. Ayurvedic therapies for thyroid dysfunction. In: Scientific Basis of Ayurvedic Therapies (Mishra L, ed.), Chapter 8. CRC Press, Boca Raton, FL, USA, 2004; pp. 133-148.
- Kar A, Panda S. Plant extracts in the regulation of hypothyroidism. In: Recent Progress in Medicinal Plants (Sharma SK, Govil JN, Singh VK, eds.), Vol. 10, Phytotherapeutics. Studium Press LLC, Houston, TX, USA, 2005; pp. 419-426.
- Carter D. British Medical Association Board of Science and Education. BMA Publications Unit, London, UK, 2004.
- Tiwari AK. Antioxidants: New generation therapeutic base for treatment of polygenic disorders. Curr Sci. 2004; 86:1092-1102.
- Chun AK. Thyroid disorders. In: Fundamentals of Geriatric Medicine (Chun AK, ed.), Chapter 25. Springer, New York, USA, 2007; pp. 450-469.
- Lindholm LH, Mendis S. Prevention of cardiovascular disease in developing countries. Lancet. 2007; 370:720-722.
- Faquin WC. The thyroid gland: Recurring problems in histologic and cytologic evaluation. Arch Pathol Lab Med. 2008; 132:622-632.
- Nattrass M, Bailey CJ. New agents for Type 2 diabetes. Baillieres Best Pract Res Clin Endocrinol Metab. 1999; 13:309-329.
- Klotz U, Sailer D. Drug interactions Their impact on safe drug therapy in the example of the new thiazolidinedione group (glitazone). Arzneimittelforschung. 2001; 51:112-117. (in German)

- Kajinami K, Takekoshi N, Saito Y. Pitavastatin: Efficacy and safety profiles of a novel synthetic HMG-CoA reductase inhibitor. Cardiovasc Drug Rev. 2003; 21:199-215.
- Cavero I, Crumb W. ICH S7B draft guideline on the nonclinical strategy for testing delayed cardiac repolarisation risk of drugs: A critical analysis. Expert Opin Drug Saf. 2005; 4:509-530.
- 14. Chatterjee S, Tringham JR, Davies MJ. Insulin glargine and its place in the treatment of Types 1 and 2 diabetes mellitus. Expert Opin Pharmacother. 2006; 7:1357-1371.
- Cabebe E, Wakelee H. Role of anti-angiogenesis agents in treating NSCLC: Focus on bevacizumab and VEGFR tyrosine kinase inhibitors. Curr Treat Options Oncol. 2007; 8:15-27.
- Cravedi JP, Zalko D, Savouret JF, Menuet A, Jégou B. The concept of endocrine disruption and human health. Med Sci (Paris). 2007; 23:198-204.
- 17. Espinosa AV, Porchia L, Ringel MD. Targeting BRAF in thyroid cancer. Br J Cancer. 2007; 96:16-20.
- Ivachtchenko AV, Kiselyov AS, Tkachenko SE, Ivanenkov YA, Balakin KV. Novel mitotic targets and their small-molecule inhibitors. Curr Cancer Drug Targets. 2007; 7:766-784.
- Nelkin BD, de Bustros AC, Mabry M, Baylin SB. The molecular biology of medullary thyroid carcinoma. A model for cancer development and progression. JAMA. 1989; 261:3130-3135.
- Gertz MA. New targets and treatments in multiple myeloma: Src family kinases as central regulators of disease progression. Leuk Lymphoma. 2008; 49:2240-2245.
- Tulis DA. Novel therapies for cyclic GMP control of vascular smooth muscle growth. Am J Ther. 2008; 15:551-564.
- Heng DY, Bukowski RM. Anti-angiogenic targets in the treatment of advanced renal cell carcinoma. Curr Cancer Drug Targets. 2008; 8:676-682.
- Mohamed Q, Wong TY. Emerging drugs for diabetic retinopathy. Expert Opin Emerg Drugs. 2008; 13:675-694.
- Rovere RK, Awada A. Treatment of recurrent thyroid cancers – is there a light in the horizon? Curr Opin Oncol. 2008; 20:245-248.
- Szczepankiewicz BG, Ng PY. Sirtuin modulators: Targets for metabolic diseases and beyond. Curr Top Med Chem. 2008; 8:1533-1544.
- Collino M, Patel NS, Thiemermann C. PPARs as new therapeutic targets for the treatment of cerebral ischemia/ reperfusion injury. Ther Adv Cardiovasc Dis. 2008; 2:179-197.
- Haverslag R, Pasterkamp G, Hoefer IE. Targeting adhesion molecules in cardiovascular disorders. Cardiovasc Hematol Disord Drug Targets. 2008; 8:252-260.
- Hermansen K, Mortensen LS, Hermansen ML. Combining insulins with oral antidiabetic agents: Affect on hyperglycemic control, markers of cardiovascular risk and disease. Vasc Health Risk Manag. 2008; 4:561-574.
- 29. Krentz AJ, Patel MB, Bailey CJ. New drugs for Type 2 diabetes mellitus: What is their place in therapy? Drugs. 2008; 68:2131-2162.
- Khama-Murad AKh, Pavlinova LI, Mokrushin AA. Hemorrhagic stroke: Molecular mechanisms of pathogenesis and perspective therapeutic targets. Usp Fiziol Nauk. 2008; 39:45-65. (in Russian)
- 31. Thomas M. Molecular targeted therapy for hepatocellular

carcinoma. J Gastroenterol. 2009; 44:136-141.

- 32. Steigen SE, Eide TJ. Gastrointestinal stromal tumors (GISTs): A review. APMIS. 2009; 117:73-86.
- Haberland M, Montgomery RL, Olson EN. The many roles of histone deacetylases in development and physiology: Implications for disease and therapy. Nat Rev Genet. 2009; 10:32-42.
- 34. Idelevich E, Kirch W, Schindler C. Current pharmacotherapeutic concepts for the treatment of obesity in adults. Ther Adv Cardiovasc Dis. 2009; 3:75-90.
- Maiese K, Chong ZZ, Shang YC, Hou J. FoxO proteins: Cunning concepts and considerations for the cardiovascular system. Clin Sci (Lond). 2009; 116:191-203.
- Singh V, Tiwari RL, Dikshit M, Barthwal MK. Models to study atherosclerosis: A mechanistic insight. Curr Vasc Pharmacol. 2009; 7:75-109.
- Sogno I, Vannini N, Lorusso G, Cammarota R, Noonan DM, Generoso L, Sporn MB, Albini A. Anti-angiogenic activity of a novel class of chemopreventive compounds: Oleanic acid terpenoids. Recent Results Cancer Res. 2009; 181:209-212.
- Katzung BG. Basic & Clinical Pharmacology. 7th ed., Applenton and Lange, Stamford, CT, USA, 1998.
- Michaelson J. Thiazolidinedione associated volume overload and pulmonary hypertension. Ther Adv Cardiovasc Dis. 2008; 2:435-438.
- Gallwitz B. Saxagliptin, a dipeptidyl peptidase IV inhibitor for the treatment of Type 2 diabetes. IDrugs. 2008; 11:906-917.
- Steinberg M. Ixabepilone: A novel microtubule inhibitor for the treatment of locally advanced or metastatic breast cancer. Clin Ther. 2008; 30:1590-1617.
- Mazzini MJ, Monahan KM. Pharmacotherapy for atrial arrhythmias: Present and future. Heart Rhythm. 2008; 5 (Suppl):S26-S31.
- Barry PJ, Gallagher P, Ryan C. Inappropriate prescribing in geriatric patients. Curr Psychiatry Rep. 2008; 10:37-43.
- Barni S, Cabiddu M, Petrelli F. Toxicity of targeted therapies in elderly patients. Expert Rev Anticancer Ther. 2008; 8:1965-1976.
- Hoffman AG, Schram SE, Ercan-Fang NG, Warshaw EM. Type I allergy to insulin: Case report and review of localized and systemic reactions to insulin. Dermatitis. 2008; 19:52-58.
- Hausner E, Fiszman ML, Hanig J, Harlow P, Zornberg G, Sobel S. Long-term consequences of drugs on the paediatric cardiovascular system. Drug Saf. 2008; 31:1083-1096.
- Jones RL. Utility of dexrazoxane for the reduction of anthracycline-induced cardiotoxicity. Expert Rev Cardiovasc Ther. 2008; 6:1311-1317.
- Nakae D, Onodera H, Fueki O, Urano T, Komiyama N, Sagami F, Kai S, Nishimura C, Inoue T. Points to consider on the non-clinical safety evaluation of anticancer drugs. J Toxicol Sci. 2008; 33:123-126.
- Pourpak Z, Fazlollahi MR, Fattahi F. Understanding adverse drug reactions and drug allergies: Principles, diagnosis and treatment aspects. Recent Pat Inflamm Allergy Drug Discov. 2008; 2:24-46.
- Yap KY, Chui WK, Chan A. Drug interactions between chemotherapeutic regimens and antiepileptics. Clin Ther. 2008; 30:1385-1407.
- Hellberg V, Wallin I, Eriksson S, Hernlund E, Jerremalm E, Berndtsson M, Eksborg S, Arnér ES, Shoshan M,

Ehrsson H, Laurell G. Cisplatin and oxaliplatin toxicity: Importance of cochlear kinetics as a determinant for ototoxicity. J Natl Cancer Inst. 2008; 101:37-47.

- Parmar HS, Kar A. Protective role of *Citrus sinensis*, *Musa paradisiaca* and *Punica granatum* peels against diet-induced atherosclerosis and thyroid dysfunctions in rats. Nutr Res. 2007; 27:710-718.
- Parmar HS, Kar A. Antidiabetic potential of *Citrus* sinensis and *Punica granatum* peel extracts in alloxan treated male mice. Biofactors. 2007; 31:17-24.
- Eddouks M, Maghrani M, Louedec L, Haloui M, Michel JB. Antihypertensive activity of the aqueous extract of *Retama raetam* Forssk. leaves in spontaneously hypertensive rats. J Herb Pharmacother. 2007; 7:65-77.
- Oliveira HC, dos Santos MP, Grigulo R, Lima LL, Martins DT, Lima JC, Stoppiglia LF, Lopes CF, Kawashita NH. Antidiabetic activity of *Vatairea macrocarpa* extract in rats. J Ethnopharmacol. 2008; 115:515-519.
- Reddy SS, Karuna R, Baskar R, Saralakumari D. Prevention of insulin resistance by ingesting aqueous extract of *Ocimum sanctum* to fructose-fed rats. Horm Metab Res. 2008; 40:44-49.
- Venables MC, Hulston CJ, Cox HR, Jeukendrup AE. Green tea extract ingestion, fat oxidation, and glucose tolerance in healthy humans. Am J Clin Nutr. 2008; 87:778-784.
- Yiming L, Wei H, Aihua L, Fandian Z. Neuroprotective effects of breviscapine against apoptosis induced by transient focal cerebral ischaemia in rats. J Pharm Pharmacol. 2008; 60:349-355.
- Agarwal A. Current issues in quality control of natural products. Pharma Times. 2005; 37:9-11.
- 60. Leontowicz M, Gorinstein S, Leontowicz H, Krzeminski R, Lojek A, Katrich E, Cíz M, Martin-Belloso O, Soliva-Fortuny R, Haruenkit R, Trakhtenberg S. Apple and pear peel and pulp and their influnce on plasma lipids an antioxidant potentials in rats fed cholesterol-containing diets. J Agric Food Chem. 2003; 51:5780-5785.
- Ma CM, Cai SQ, Cui JR, Wang RQ, Tu PF, Hattori M, Daneshtalab M. The cytotoxic activity of ursolic acid derivatives. Eur J Med Chem. 2005; 40:582-589.
- Dixit Y, Panda S, Kar A. *Lagenaria siceraria* peel extract in the regulation of hyperthyroidism, hyperglycemia and lipid peroxidation in mice. Int J Biomed Pharma Sci. 2008; 2:79-83.
- Parmar HS, Kar A. Protective role of *Mangifera indica*, *Cucumis melo* and *Citrullus vulgaris* peel extracts in chemically induced hypothyroidism. Chem Biol Interac. 2009; 177:254-258.
- Parmar HS, Kar A. Comparative analysis of free radical scavenging potential of several fruit peel extracts by *in vitro* methods. Drug Disc Ther. 2009; 3:49-55.
- 65. Parmar HS, Kar A. Medicinal values of fruit peels from *Citrus sinensis, Punica granatum* and *Musa paradisiaca* with respect to alterations in tissue lipid peroxidation and serum concentration of glucose, insulin and thyroid hormones. J Med Food. 2008; 11:376-381.
- Parmar HS, Kar A. Antiperoxidative, antithyroidal, antihyperglycemic and cardioprotective role of *Citrus sinensis* peel extracts in male mice. Phytother Res. 2008; 22:791-795.
- 67. Parmar HS, Kar A. Possible amelioration of atherogenic diet induced dyslipidemia, hypothyroidism and hyperglycemia by the peel extracts of *Mangifera indica*, *Cucumis melo* and *Citrullus vulgaris* fruits in rats.

Biofactors. 2008; 33:13-24.

- Rincón AM, Vásquez AM, Padilla FC. Chemical composition and bioactive compounds of flour of orange (*Citrus sinensis*), tangerine (*Citrus reticulata*) and grapefruit (*Citrus paradisi*) peels cultivated in Venezuela. Arch Latinoam Nutr. 2005; 55:305-310. (in Spanish)
- Ho SC, Lin CC. Investigation of heat treating conditions for enhancing the anti-inflammatory activity of citrus fruit (*Citrus reticulata*) peels. J Agric Food Chem. 2008; 56:7976-7982.
- Tommonaro G, Rodríguez CS, Santillana M, Immirzi B, Prisco RD, Nicolaus B, Poli A. Chemical composition and biotechnological properties of a polysaccharide from the peels and antioxidative content from the pulp of *Passiflora liguralis* fruits. J Agric Food Chem. 2007; 55:7427-7433.
- Leontowicz H, Gorinstein S, Lojek A, Leontowicz M, Cíz M, Soliva-Fortuny R, Park YS, Jung ST, Trakhtenberg S, Martin-Belloso O. Comparative content of some bioactive compounds in apples, peaches and pears and their influence on lipids and antioxidants capacity in rats. J Nutr Biochem. 2002; 13:603-610.
- Negro C, Tommasi L, Miceli A. Phenolic compounds and antioxidant from red grape marc extracts. Bioresour Technol. 2003; 87:41-44.
- Noda Y, Kaneyuki T, Igarashi K, Mori A, Packer L. Antioxidant activity of nasunin, an anthocyanin in eggplant. Res Commun Mol Pathol Pharmacol. 1998; 102:175-187.
- Sadilova E, Stintzing FC, Carle R. Anthocyanins, colour and antioxidant properties of eggplant (*Solanum melongena* L.) and violet pepper (*Capsicum annuum* L.) peel extracts. Z Naturforsch C. 2006; 61:527-535.
- Singh N, Kamath V, Rajini PS. Protective effect of potato peel power in ameliorating oxidative stress in streptozotocin diabetic rats. Plant Foods Hum Nutr. 2005; 60:49-54.
- Singh N, Rajini PS. Antioxidant-mediated protective effect of potato peel extract in erythrocytes against oxidative damage. Chem Biol Interact. 2008; 173:97-104.
- 77. Gorinstein S, Leontowicz H, Leontowicz M, Drzewiecki J, Jastrzebski Z, Tapia MS, Katrich E, Trakhtenberg S. Red Star Ruby (Sunrise) and blond qualities of Jaffa grapefruits and their influence on plasma lipid levels and plasma antioxidant activity in rats fed with cholesterol-containing and cholesterol-free diets. Life Sci. 2005; 77:2384-2397.
- Matsubara Y, Kumamoto H, Iizuka Y, Murakami T, Okamoto K, Miyake H, Yokoi K. Structure and hypertensive effect of flavonoids glycosides in *Citrus unshiu* peelings. Agric Biol Chem. 1985; 49:909-914.
- Narayana RK, Reddy MS, Chaluvadi MR, Krishna DR. Bioflavonids classification, pharmacological, biochemical effects and therapeutic potential. Ind J Pharmacol. 2001; 33:2-16.
- Jung UJ, Lee MK, Jeong KS, Choi MS. The hypoglycemic effects of hesperidin and naringin are partly mediated by hepatic glucose-regulating enzymes in C57BL/KsJ-db/db mice. J Nutr. 2004; 134:2499-2503.
- Fuhrman B, Lavy A, Aviram M. Consumption of red wine with meals reduces the susceptibility of human plasma and low-density lipoproteins to lipid peroxidation. Am J Clin Nutr. 1995; 61:549-554.
- 82. Igarashi K, Ohmuna M. Effect of Isoharnneti, Rhamnetin and Quercetin on the concentrations of cholesterol and lipoperoxide in the serum and liver and on the blood

and liver antioxidative enzyme activities in rats. Biosci Biotech Biochem. 1995; 59:595-601.

- McAnlis GT, McEneny J, Pearce J, Young IS. The effect of various dietary flavonoids on the susceptibility of low density lipoproteins to oxidation *in vitro* using both metallic and non-metallic oxidising agents. Biochem Soc Trans. 1997; 25:142S.
- 84. Bok SH, Lee SH, Park YB, Bae KH, Son KH, Jeong TS, Choi MS. Plasma and hepatic cholesterol and hepatic activities of 3-hydroxyl-3-methyl-glutaryl-CoA reductase and acyl CoA: Cholesterol transferase are lower in rats fed citrus peel extract or a mixture of citrus bioflavonoids. J Nutr. 1999; 129:1182-1185.
- Choi MS, Do KM, Park YS, Jeon SM, Jeong TS, Lee YK, Lee MK, Bok SH. Effect of naringin supplementation on cholesterol metabolism and antioxidant status in rats fed high cholesterol with different levels of vitamin E. Ann Nutr Metab. 2001; 45:193-201.
- de Whalley CV, Rankin SM, Hoult JR, Jessup W, Leake DS. Flavonoids inhibit the oxidative modification of low density lipoproteins by macrophages. Biochem Pharmacol. 1990; 39:1743-1750.
- Francel EN, Kanner J, German JB, Parks E, Kinsella JE. Inhibition of oxidation of human low-density lipoprotein by phenolis substances in red wine. Lancet. 1993; 341:454-457.
- Kroyer G. The antioxidant activity of citrus fruit peels. Z Ernahrungswiss. 1986; 25:63-69. (in German)
- Böhm H, Boeing H, Hempel J, Raab B, Kroke A. Flavonols, flavone and anthocyanins as natural antioxidants of food and their possible role in the prevention of chronic diseases. Z Ernahrungswiss. 1998; 37:147-163. (in German)
- 90. Murakami A, Nakamura Y, Ohto Y, Yano M, Koshiba T, Koshimizu K, Tokuda H, Nishino H, Ohishi H. Suppressive effects of citrus fruits on free radical generation and nobiletin, an anti-inflammatory polymethoxylated flavonoid. Biofactors. 2000; 12:187-192.
- Choe SC, Kim HS, Jeong TS, Bok SH, Park YB. Naringin has an antiatherogenic effect with the inhibition of intercellular adhesion molecule-1 in hypercholesterolemic rabbits. J Cardiovasc Pharmacol. 2001; 38:947-955.
- 92. Anagnostopoulou MA, Kefalas P, Kokkalou E, Assimopoulou AN, Papageorgiou VP. Analysis of antioxidant compounds in sweet orange peel by HPLCdiode array detection-electrospray ionization mass spectrometry. Biomed Chromatogr. 2005; 19:138-148.
- Lyte M. Induction of gram-negative bacterial growth by neurochemical containing banana (*Musa x paradisiaca*) extracts. FEMS Microbiol Lett. 1997; 154:245-250.
- Kanazawa K, Sakakibara H. High content of dopamine, a strong antioxidant, in Cavendish banana. J Agric Food Chem. 2000; 48:844-848.
- 95. Deepa PR, Varalakshmi P. Salubrious effect of low molecular weight heparin on atherogenic diet-induced cardiac, hepatic and renal lipid peroxidation and collapse of antioxidant defences. Mol Cell Biochem. 2003; 254:111-116.
- Deepa PR, Varlakshmi P. Protective effects of certoparin sodium, a low molecular weight heparin derivative in experimental atherosclerosis. Clin Chim Acta. 2004; 339:105-115.
- 97. Jatwa R, Kar A. Cardio-protective role of terazosin is possibly mediated through alteration in thyroid function.

Eur J Pharmacol. 2006; 551:87-91.

- Vedavanam K, Srijayanta S, O'Reilly J, Raman A, Wiseman H. Antioxidant action and potential antidiabetic properties of an isoflavonoid-containing soyabean phytochemical extract (SPE). Phytother Res. 1999; 13:601-608.
- Murthy KN, Reddy VK, Veigas JM, Murthy UD. Study on wound healing activity of *Punica granatum* peel. J Med Food. 2004; 7:256-259.
- 100. Seeram NP, Adams LS, Henning SM, Niu Y, Zhang Y, Nair MG, Heber D. *In vitro* antiproliferative, apoptotic and antioxidant activities of punicalagin, ellagic acid and a total pomegranate tannin extract are enhanced in combination with other polyphenols as found in pomegranate juice. J Nutr Biochem. 2005; 16:360-367.
- 101. Cazarolli LH, Zanatta L, Jorge AP, de Sousa E, Horst H, Woehl VM, Pizzolatti MG, Szpoganicz B, Silva FR. Follow-up studies on glycosylated flavonoids and their complexes with vanadium: Their anti-hyperglycemic potential role in diabetes. Chem Biol Interact. 2006; 163:177-191.
- 102. Aslan M, Deliorman Orhan D, Orhan N, Sezik E, Yesilada E. *In vivo* antidiabetic and antioxidant potential of *Helichrysum plicatum* ssp. *plicatum* capitulums in streptozotocin-induced-diabetic rats. J Ethnopharmacol. 2007; 109:54-59.
- 103. Chemler JA, Lock LT, Koffas MA, Tzanakakis ES. Standardized biosynthesis of flavan-3-ols with effects on pancreatic beta-cell insulin secretion. Appl Microbiol Biotechnol. 2007; 77:797-807.
- 104. Hsu CL, Yen GC. Effect of gallic acid on high fat dietinduced dyslipidaemia, hepatosteatosis and oxidative stress in rats. Br J Nutr. 2007; 98:727-735.
- 105. Katz SR, Newman RA, Lansky EP. Punica granatum L.: Heuristic treatment for diabetes mellitus. J Med Food. 2007; 10:213-217.
- 106. Jang A, Srinivasan P, Lee NY, Song HP, Lee JW, Lee M, Jo C. Comparison of hypolipidemic activity of synthetic gallic acid-linoleic acid ester with mixture of gallic acid and linoleic acid, gallic acid, and linoleic acid on highfat diet induced obesity in C57BL/6 Cr Slc mice. Chem Biol Interact. 2008; 174:109-117.
- 107. Kato A, Nasu N, Takebayashi K, Adachi I, Minami Y, Sanae F, Asano N, Watson AA, Nash RJ. Structureactivity relationships of flavonoids as potential inhibitors of glycogen phosphorylase. J Agric Food Chem. 2008; 56:4469-4473.
- 108. Mitchell DE, Madore MA. Patterns of assimilate production and translocation in Muskmelon (*Cucumis melo* L.): II. Low temperature effects. Plant Physiol. 1992; 99:966-971.
- 109. Ganong W. Review of Medical Physiology. 17th ed., Applenton and Lange, Stamford, CT, USA, 2005.
- 110. Percival SS, Talcott ST, Chin ST, Mallak AC, Lounds-Singleton A, Pettit-Moore J. Neoplastic transformation of BALB/3T3 cells and cell cycle of HL-60 cells are inhibited by mango (*Mangifera indica* L.) juice and mango juice extracts. J Nutr. 2006; 136:1300-1304.
- 111. Rodríguez J, Di Peirro D, Gioia M, Monaco S, Delgado R, Coletta M, Marini S. Effects of a natural extract from *Mangifera indica* L., and its active compound, mangiferin, on energy state and lipid peroxidation of red blood cells. Biochim Biophys Acta. 2006; 1760:1333-1342.
- 112. Sa-Nunnes A, Rogerio AP, Medeiros AI, Fabris VE,

Andreu GP, Rivera DG, Delgado R, Faccioli LH. Modulation of eosinophil generation and migration by *Mangifera indica* L. extract (Vimang). Int Immunopharmacol. 2006; 6:1515-1523.

- 113. Hernandez P, Rodriguez PC, Delgado R, Walczak H. Protective effect of *Mangifera indica* L. polyphenols on human T lymphocytes against activation-induced cell death. Pharmacol Res. 2007; 55:167-173.
- 114. Selles AJ, Rodriguez MD, Balseiro ER, Gonzalez LN, Nicolais V, Rastrelli L. Comparison of major and trace element concentrations in 16 varieties of Cuban mango stem bark (*Mangifera indica* L.). J Agric Food Chem. 2007; 55:2176-2181.
- 115. Knödler M, Conrad J, Wenzig EM, Bauer R, Lacorn M, Beifuss U, Carle R, Schieber A. Anti-inflammatory 5-(11'Z-heptadecenyl)-and 5-(8'Z,11'Z-heptadecadienyl)resorcinols from mango (*Mangifera indica* L.) peels. Phytochem. 2008; 69:988-993.
- 116. Castillo L, Sanchez M, Vogt J, Chapman TE, De Rojas-Walker TC, Tannenbaun SR, Ajami AM, Young VR. Plasma arginine, citrulline, and ornithine kinetics in adults, with observations on nitric oxide synthesis. Am J Physiol. 1995; 268:E360-E367.
- 117. Edwards AJ, Vinyard BT, Wiley ER, Brown ED, Collins JK, Perkins-Veazie P, Baker RA, Clevidence BA. Consumption of watermelon juice increases plasma concentrations of lycopene and beta-carotene in humans. J Nutr. 2003; 133:1043-1050.
- 118. Collins JK, Wu G, Perkins-Veazie P, Spears K, Claypool PL, Baker RA, Clevidence BA. Watermelon consumption increases plasma arginine concentration in adults. Nutrition. 2007; 23:261-266.
- 119. Chau CF, Huang YL, Lee MH. *In vitro* hypoglycemic effects of different insoluble fiber-rich fractions prepared from the peel of *Citrus sinensis* L. cv, Liucheng. J Agric Food Chem. 2003; 51:6623-6626.
- 120. Auf'mkolk M, Köhrle J, Gumbinger H, Winterhoff H, Hesch RD. Antihormonal effects of plant extracts: Iodothyronine deiodinase of rat liver is inhibited by extracts and secondary metabolites of plants. Horm Metab Res. 1984; 16:188-192.
- 121. Piattelli M, Impellizzeri G. Fungistatic flavones in the leaves of citrus species resistant and susceptible to *deuterophoma tracheiphila*. Phytochemistry. 1971; 10:2657-2659.
- 122. Tatum JH, Berry RE. Six new flavonoids from *Citrus*. Phytochemistry. 1972; 11:2283-2288.
- 123. Divi RL, Doerge DR. Inhibition of thyroid peroxidase by dietary flavonoids. Chem Res Toxicol. 1996; 9:16-23.
- Wilding J, Williams G. Textbook of Medicine. 3rd ed., Churchill Livingstone, London, UK, 1997.
- 125. Filippi L, Cecchi A, Tronchin M, Dani C, Pezzati M, Seminara S, Gasperini S, Zammarchi E, Rubaltelli FF. Dopamine infusion and hypothyroxinaemia in very low birth weight preterm infants. Eur J Pediatr. 2004; 163:7-13.
- 126. da-Silva WS, Harney JW, Kim BW, Li J, Bianco SD, Crescenzi A, Christoffolete MA, Huang SA, Bianco AC. The small polyphenolic molecule kaempferol increases cellular energy expenditure and thyroid hormone activation. Diabetes. 2007; 56:767-776.
- 127. Wolfe KL, Liu RH. Apple peels as a value-added food ingredient. J Agric Food Chem. 2003; 51:1676-1683.
- 128. Yance DR Jr, Sagar SM. Targeting angiogenesis with integrative cancer therapies. Integr Cancer Ther. 2006;

325

5:9-29.

- 129. Mahadevan S, Park Y. Multifaceted therapeutic benefits of *Ginkgo biloba* L.: Chemistry, efficacy, safety, and uses. J Food Sci. 2008; 73:R14-R19.
- 130. He X, Liu RH. Phytochemicals of apple peels: Isolation, structure elucidation, and their antiproliferative and antioxidant activities. J Agric Food Chem. 2008; 56:9905-9910.
- 131. Yoon H, Liu RH. Effect of 2alpha-hydroxyursolic acid on NF-kappaB activation induced by TNF-alpha in human breast cancer MCF-7 cells. J Agric Food Chem. 2008; 56:8412-8417.
- 132. Li-Weber M. New therapeutic aspects of flavones: The anticancer properties of *Scutellaria* and its main active constituents *wogonin*, *baicalein* and *baicalin*. Cancer Treat Rev. 2009; 35:57-68.
- 133. Chinnici F, Bendini A, Gaiani A, Riponi C. Radical scavenging activities of peels and pulps from cv. Golden Delicious apples as related to their phenolic composition. J Agric Food Chem. 2004; 52:4684-4689.
- 134. He X, Liu RH. Triterpenoids isolated from apple peels have potent antiproliferative activity and may be partially responsible for apple's anticancer activity. J Agric Food Chem. 2007; 55:4366-4370.
- 135. Lansky EP, Newman RA. Punica granatum (pomegranate) and its potential for prevention and treatment of inflammation and cancer. J Ethnopharmacol. 2007; 109:177-206.
- 136. Murakami A, Kuki W, Takahashi Y, Yonei H, Nakamura Y, Ohto Y, Ohigashi H, Koshimizu K. Auraptene, a citrus coumarin, inhibits 12-O-tetradecanoylphorbol-13acetate-induced tumor promotion in ICR mouse skin, possibly through suppression of superoxide generation in leukocytes. Jpn J Cancer Res. 1997; 88:443-452.
- 137. Kadota Y, Taniguchi C, Masuhara S, Yamamoto S, Furusaki S, Iwahara M, Goto K, Matsumoto Y, Ueoka R. Inhibitory effects of extracts from peels of *Citrus natsudaidai* encapsulated in hybrid liposomes on the growth of tumor cells *in vitro*. Biol Pharm Bull. 2004; 27:1465-1467.
- 138. Kim MJ, Park HJ, Hong MS, Park HJ, Kim MS, Leem KH, Kim JB, Kim YJ, Kim HK. Citrus reticulate blanco induces apoptosis in human gastric cancer cells SNU-668. Nutr Cancer. 2005; 51:78-82.
- 139. Iwase Y, Takemura Y, Ju-ichi M, Kawaii S, Yano M, Okuda Y, Mukainaka T, Tsuruta A, Okuda M, Takayasu J, Tokuda H, Nishino H. Inhibitory effect of Epstein-Barr virus activation by *Citrus fruits*, a cancer chemopreventor. Cancer Lett. 1999; 139:227-236.
- 140. Kawase M, Motohashi N, Satoh K, Sakagami H, Nakashima H, Tani S, Shirataki Y, Kurihara T, Spengler G, Wolfard K, Molnár J. Biological activity of persimmon (*Diospyros kaki*) peel extracts. Phytother Res. 2003; 17:495-500.
- 141. Motohashi N, Kawase M, Shirataki Y, Tani S, Saito S, Sakagami H, Kurihara T, Nakashima H, Wolfard K, Mucsi I, Varga A, Molnár J. Biological activity of feijoa peel extracts. Anticancer Res. 2000; 20:4323-4329.
- 142. Azuma K, Ohyama A, Ippoushi K, Ichiyanagi T, Takeuchi A, Saito T, Fukuoka H. Structures and antioxidant activity of anthocyanins in many accessions of eggplant and its related species. J Agric Food Chem. 2008; 56:10154-10159.
- 143. Gorinstein S, Zachwieja Z, Folta M, Barton H, Piotrowicz J, Zemser M, Weisz M, Trakhtenberg S,

Màrtín-Belloso O. Comparative contents of dietary fiber, total phenolics, and minerals in persimmons and apples. J Agric Food Chem. 2001; 49:952-957.

- 144. Kawaguchi K, Mizuno T, Aida K, Uchino K. Hesperidin as an inhibitor of lipases from porcine pancreas and Pseudomonas. Biosci Biotechnol Biochem. 1997; 61:102-104.
- 145. Lai CS, Li S, Chai CY, Lo CY, Ho CT, Wang YJ, Pan MH. Inhibitory effect of citrus 5-hydroxy-3,6,7,8,3',4'hexamethoxyflavone on 12-O-tetradecanoylphorbol 13-acetate-induced skin inflammation and tumor promotion in mice. Carcinogenesis. 2007; 28:2581-2588.
- 146. Morita H, Enomoto M, Hirasawa Y, Iizuka T, Ogawa K, Kawahara N, Goda Y, Matsumoto T, Takeya K. Cyclonatsudamine A, a new vasodilator cyclic peptide from *Citrus natsudaidai*. Bioorg Med Chem Lett. 2007; 17:5410-5413.
- 147. Ma YQ, Ye XQ, Fang ZX, Chen JC, Xu GH, Liu DH. Phenolic compounds and antioxidant activities of extracts from ultrasonic treatment of Satsuma Mandarin (*Citrus unshiu* Marc.) peels. J Agric Food Chem. 2008; 56:5682-5690.
- 148. Xu GH, Chen JC, Liu DH, Zhang YH, Jiang P, Ye XQ. Minerals, phenolic compounds, and antioxidant capacity of citrus peel extract by hot water. J Food Sci. 2008; 73: C11-C18.
- 149. Choi SY, Ko HC, Ko SY, Hwang JH, Park JG, Kang SH, Han SH, Yun SH, Kim SJ. Correlation between flavonoid content and the NO production inhibitory activity of peel extracts from various citrus fruits. Biol Pharm Bull. 2007; 30:772-778.
- 150. Schieber A, Berardini N, Carle R. Identification of flavonol and xanthone glycosides from mango (*Mangifera indica* L., Cv. "TommyAtkins") peels by high-performanceliquid chromatography-electrospray ionization mass spectrometry. J Agric Food Chem. 2003; 51:5006-5011.
- 151. Barreto JC, Trevisan MT, Hull WE, Erben G, de Brito ES, Pfundstein B, Würtele G, Spiegelhalder B, Owen RW. Characterization and quantitation of polyphenolic compounds in bark, kernel, leaves, and peel of mango (*Mangifera indica* L.). J Agric Food Chem. 2008; 56:5599-5610.
- 152. Knödler M, Conrad J, Wenzig EM, Bauer R, Lacorn M, Beifuss U, Carle R, Schieber A. Anti-inflammatory 5-(11'Z-heptadecenyl)- and 5-(8'Z,11'Z-heptadecadienyl)-resorcinols from mango (*Mangifera indica* L.) peels. Phytochemistry. 2008; 69:988-993.
- 153. Tommonaro G, Rodriguez CS, Santillana M, Immirzi B, Prisco RD, Nicolaus B, Poli A. Chemical composition and biotechnological properties of a polysaccharide from the peels and antioxidative content from the pulp of *Passiflora liquralis* fruits. J Agric Food Chem. 2007; 55:7427-7433.
- 154. Fiorentino A, D'Abrosca B, Pacifico S, Mastellone C, Piscopo V, Caputo R, Monaco P. Isolation and structure elucidation of antioxidant polyphenols from quince (*Cydonia vulgaris*) peels. J Agric Food Chem. 2008; 56:2660-2667.
- 155. Reboul E, Borel P, Mikail C, Abou L, Charbonnier M, Caris-Veyrat C, Goupy P, Portugal H, Lairon D, Amiot MJ. Enrichment of tomato paste with 6% tomato peel increases lycopene and beta-carotene bioavailability in men. J Nutr. 2005; 135:790-794.

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