

**ScienceDirect** 



# **Transmission routes of respiratory viruses among humans** Jasmin S Kutter<sup>1,3</sup>, Monique I Spronken<sup>1,3</sup>, Pieter L Fraaij<sup>1,2</sup>,



Respiratory tract infections can be caused by a wide variety of viruses. Airborne transmission via droplets and aerosols enables some of these viruses to spread efficiently among humans, causing outbreaks that are difficult to control. Many outbreaks have been investigated retrospectively to study the possible routes of inter-human virus transmission. The results of these studies are often inconclusive and at the same time data from controlled experiments is sparse. Therefore, fundamental knowledge on transmission routes that could be used to improve intervention strategies is still missing. We here present an overview of the available data from experimental and observational studies on the transmission routes of respiratory viruses between humans, identify knowledge gaps, and discuss how the available knowledge is currently implemented in isolation guidelines in health care settings.

Ron AM Fouchier<sup>1</sup> and Sander Herfst<sup>1</sup>

#### Addresses

<sup>1</sup> Department of Viroscience, Postgraduate School of Molecular Medicine, Erasmus Medical Centre, Rotterdam, The Netherlands <sup>2</sup> Department of Pediatrics, Subdivision Infectious diseases and Immunology, Erasmus Medical Centre – Sophia, Rotterdam, The Netherlands

Corresponding author: Herfst, Sander (s.herfst@erasmusmc.nl) <sup>3</sup>These authors contributed equally to this work.

#### Current Opinion in Virology 2018, 28:142-151

This review comes from a themed issue on **Emerging viruses:** intraspecies transmission

Edited by Sander Herfst and Martin Ludlow

For a complete overview see the Issue and the Editorial

Available online 17th January 2018

https://doi.org/10.1016/j.coviro.2018.01.001

1879-6257/© 2018 The Authors. Published by Elsevier B.V. This is an open access article under the CC BY-NC-ND license (http://creative-commons.org/licenses/by-nc-nd/4.0/).

### Introduction

Viral respiratory tract infections are a leading cause of morbidity and mortality worldwide, representing an enormous economic and disease burden [1]. Respiratory viruses replicate in the respiratory tract from where they are subsequently shed and transmitted via respiratory secretions. They are classified in different virus families and differ in virulence and target groups. Respiratory tract infections may range from asymptomatic to acute live threating disease thereby posing a major health threat to young children, elderly, and immunocompromised people. Respiratory viruses spread via three different transmission routes: contact (direct or indirect), droplet and aerosol transmission (Table 1) [2,3]. Contact transmission refers to direct virus transfer from an infected person to a susceptible individual (e.g. via contaminated hands) or indirect virus transfer via intermediate objects (fomites). Transmission of virus through the air can occur via droplets or aerosols. The commonly accepted cut-off size between the large droplets and small aerosols is 5 µm, although this varies considerably between studies, ranging up to 12 µm [4–8]. Droplets generated during coughing, sneezing or talking do not remain suspended in air and travel less than 1 m before settling on the mucosa of close contacts or environmental surfaces. Aerosols have a slow settling velocity, thus they remain suspended in the air longer and can travel further [5,9,10].

Transmission via each of these three routes is complex and depends on many variables such as environmental factors (e.g. humidity and temperature), crowding of people, but also on host factors such as receptor distribution throughout the respiratory tract. The fact that all these variables affect the different transmission routes of the different respiratory viruses in a dissimilar way, makes it very difficult to investigate them experimentally [9,11]. Here, we summarize the evidence from experimental and observational studies on inter-human transmission routes of important respiratory viruses (summarized in Table 2). A literature search was conducted for each respiratory virus using 'human transmission experiments' and 'transmission (routes)' of the virus of interest as search criteria in PubMed and Google Scholar. Subsequently, the backward snowball method was applied in which additional papers were identified based on the reference list of a paper of interest. As this review focuses on the evidence on inter-human transmission routes, data from animal studies were excluded. In addition, intervention studies, (aircraft) outbreak reports and household studies were excluded if the transmission route was not specifically investigated. The strengths and weaknesses of the different methods employed in transmission studies are summarized in Table 3. Finally, we discuss our findings in the light of several available (inter)national guidelines on infection control. Our observations underscore the urgent need for new knowledge on respiratory virus transmission routes and the implementation of this knowledge in infection control guidelines to advance intervention strategies for currently circulating and newly emerging viruses and to improve public health.

Table 1	able 1						
Commonly accepted respiratory routes of transmission							
Transmission route	Particles involved and particle characteristics	Characteristics/definition of transmission					
Contact		Self-inoculation of mucous membranes by contaminated hands.					
Direct	Deposited on persons.	Virus transfer from one infected person to another.					
Indirect	Deposited on objects.	Virus transfer through contaminated intermediate objects (fomites)					
Airborne							
Droplet	Droplets (>5 μm).	Short range transmission.					
	Remain only shortly in air (<17 min) [116].	Direct inoculation of naïve person through coughing/sneezing/					
	Dispersed over short distances (<1 m).	breathing of infected person.					
		Deposition mainly on mucous membranes and upper respiratory					
		tract.					
Aerosol	Aerosols, droplet nuclei (<5 μm),	Long range transmission.					
	Remain in air for an almost infinite amount of time.	Inhalation of aerosols in respirable size range.					
	Dispersed over long distances (>1 m).	Deposition along the respiratory tract, including the lower airways.					

Table 2

Overview of the evidence on transmission routes of respiratory viruses based on experimental data and the transmission route according to infection prevention guidelines

Virus	Virus family <sup>a</sup>	Transmission route	
		Experimental and observational data	Guidelines <sup>b</sup>
Measles virus	Paramyxoviridae	Aerosol [75-77,78°,79°].	Contact [3,110], droplet [3,109–111], aerosol [3,109–111].
Parainfluenza virus	Paramyxoviridae	Limited data, contact (by fomite) [83,84] <sup>e</sup> .	Contact [3,109–111], droplet [3,109–111], aerosol [3,109].
HMPV	Pneumoviridae	Limited data, contact (by fomite) <sup>e</sup> [30]	Contact [3,110,111], droplet [3,110,111].
RSV	Pneumoviridae	Contact [89,88], droplet [88], aerosol [90,91**].	Contact [3,109–111], droplet [3,109,110], aerosol [109,111].
HCoV	Coronaviridae	Limited data, contact (by fomite) [65-67] e.	Contact [3,110,111], droplet [3,110,111].
MERS-CoV	Coronaviridae	Contact [84] <sup>e</sup> [89] <sup>c</sup> [91 <sup>••</sup> ], droplet [89] <sup>c</sup> , aerosol [91 <sup>••</sup> ].	Contact [111], droplet [3,111]
SARS-CoV	Coronaviridae	Contact [70] <sup>e</sup> [73,79,101], droplet [73,78 <sup>•</sup> ,79 <sup>•</sup> ,117], aerosol [76,118] <sup>c</sup> [82] <sup>c,d</sup> .	Contact [3,110,111], droplet [3,110,111], aerosol [3,110,111].
Rhinovirus	Picornaviridae	Contact [35,36,42], aerosol [37,40,119].	Contact [109–111], droplet [109,111], aerosol [109–111].
Adenovirus	Adenoviridae	Contact [100] <sup>e</sup> [100,101], droplet [103*], aerosol [102,103*].	Contact [3,109–111], droplet [3,109,110], aerosol [110,111].
Influenza virus	Orthomyxoviridae	Droplet/aerosol [55,56,57*,59]	Contact [109–111], droplet [3,109–111], aerosol [3,109–111].

<sup>a</sup> Taxonomy was based on [62], airborne transmission is seemingly linked to:

<sup>b</sup> WIP [108], 'Blue Book' [109], 'Red Book' [110], CDC [3] and Up-To-Date [111]. The conclusions on experimental data as presented in this table reflect the conclusions from the authors.

<sup>c</sup> Superspreader events.

<sup>d</sup> Aerosol-generating procedures (in a nosocomial situation).

<sup>e</sup> Conclusions were drawn based on stability experiments.

#### Measles virus (MV)

Measles is one of the most contagious viral diseases in humans that has been associated with aerosol transmission for a long time [12,13,14<sup>••</sup>,15–17,18<sup>••</sup>]. However, it should be noted that MV also replicates systemically, and that there is a role for dead cell debris-associated virus spread via fomites. In the late 1970s and early 1980s, data from retrospective observational studies obtained during outbreaks in pediatric practices, a school, and a sporting event suggested transmission through aerosols [14<sup>••</sup>,15– 17,18<sup>••</sup>]. Indeed, those studies showed that most secondary cases never came in direct contact with the index patient and some were never even simultaneously present in the same area as the index case [14\*\*,18\*\*]. Examination of airflow in the pediatricians' offices showed that aerosols were not only dispersed over the entire examination room but also accumulated in the hallway and other areas [14\*\*,18\*\*]. Furthermore, based on the investigation of air circulation in a sport stadium, in which a MV outbreak occurred, authors suggested that MV had been dispersed through the ventilation system [16]. Thus it was concluded that MV can be transmitted via aerosols. Although coughing is a common symptom associated with measles disease, index patients were described to cough

Table 3

Study design	Pro	Con	Reference
Virus stability	Can provide indirect evidence for	Not conclusive as transmission itself is not	[42,43,65,70]
Virus stability	transmission route.	investigated.	[+2,+0,00,70]
Outbreak (being bala	<ul> <li>Easy to perform.</li> <li>Study natural infections.</li> </ul>	Retrospective.	[120 122]
Outbreak (household or hospital) reports	<ul> <li>Includes the most susceptible</li> </ul>	<ul> <li>Usually not conclusive on transmission route or</li> </ul>	[120–123]
	patients who are difficult to include in experimental studies.	relative importance of transmission routes.	
Outbreak report — aircraft	Relatively easy to perform	Retrospective which can result in recall-bias and	[118,124–127]
	Outbreak in closed setting	hard to trace back passenger movements. • Inconclusive.	[,]
		Only reported in case of secondary infections and	
		in these cases infections may also occur before or	
		after the flight.	
Non-pharmaceutical	Can help to discriminate between	Usually no controlled environment.	[35,128–131]
Intervention	transmission routes if performed	• Difficult to determine ideal time-point of the	
	properly.	intervention.	
		<ul> <li>Risk of drop-out or perseverance.</li> </ul>	
Pharmaceutical	Can help to identify relative	Difficult to include enough patients to obtain	[132]
intervention	<ul><li>importance of transmission routes</li><li>Controlled environment</li></ul>	statistically significant results	
Experimental infection	<ul> <li>Controlled environment.</li> </ul>	Ethical obstacles.	[42,44,102]
	Donor selection and control.	<ul> <li>Infectivity and disease can differ from that in a</li> </ul>	
	Real-time data collection.	natural infection (attenuated strains).	
	Repeatable.	Difficult to create ideal and comparable	
	<ul> <li>Various parameters can be studied at the same time.</li> </ul>	<ul><li>circumstances.</li><li>Many factors have to be taken into account:</li></ul>	
	<ul> <li>Possibility to study different</li> </ul>	duration, influence of superspreaders, sampling	
	inoculation routes.	methods.	
		Difficult to get naïve or risk group participants	
		who are interesting to study.	
Miniature field trial	Can discriminate between contact	Ethical obstacles.	[38**,39**,40]
	and airborne transmission.	<ul> <li>Exposure time may not be sufficient.</li> </ul>	
		<ul> <li>Difficult to create ideal and comparable</li> </ul>	
		circumstances.	
Air sampling	<ul> <li>Noninvasive for patients.</li> </ul>	<ul> <li>In a nosocomial setting aerosol-generating</li> </ul>	
	Quantification of viable virus in the	procedures can play a major role.	
	air.	Frequently only detection by PCR.	
	Characterization of droplet/	Direct human-to-human transmission is not	
	<ul><li>aerosol size.</li><li>Can be used in parallel with human</li></ul>	studied (circumstantial).	
	studies or outbreaks.	• Technical issues (procedure may affect virus viability) or false interpretation.	
	Can gain information on possible	viability) of faise interpretation.	
	aerosol spread.		
	[34 <sup>••</sup> ,37,57 <sup>•</sup> ,91 <sup>••</sup> ,103 <sup>•</sup> ,133]		
Air tracer studies	<ul> <li>Monitoring airflow pattern can</li> </ul>	Usually performed retrospectively and not during	[134,135]
	indicate possible airborne	outbreaks	
	transmission (if not done		
	retrospectively).		
	Visualize airstream		
Computational	Describes transmission in a	Theoretical (for mathematical modeling).	[82,136–141]
Modeling/Simulation	greater context.	Artificial setting.	
	Can account for heterogeneity of		
	transmission within a population.		
	<ul> <li>Human mannequins can be used as replacement for humans</li> </ul>		
	as replacement for numans		

frequently and vigorously in the outbreak reports of pediatric practices. Remington *et al.* calculated the infectious dose of MV produced by the index case through coughing, using a mathematical model based on airborne transmission. They found that the index case produced a very high infectious dose compared to cases from other outbreaks and mentioned a phenomenon called superspreading [18<sup>••</sup>]. Superspreaders are individuals who are able to infect a disproportionally large number of susceptible contacts when compared to a typical individual [19– 22], which may contribute to the efficient transmission of MV.

# Parainfluenza (PIV) and human metapneumovirus (HMPV)

There is a substantial lack of (experimental) evidence on the transmission routes of PIV (types 1–4) and HMPV. For both viruses, contact and droplet transmission are commonly accepted transmission routes [23–25]. However, only virus stability on various surfaces has been investigated so far and it has been shown that PIV and HMPV are stable on non-absorptive surfaces and can barely be recovered from absorptive surfaces [26–30].

#### **Respiratory syncytial virus (RSV)**

Transmission of RSV among humans is thought to occur via droplets and fomites [1,7]. In the 1980s three potential transmission routes of RSV were studied in humans by dividing infected infants and healthy volunteers into three groups, representing: Firstly, all transmission routes, secondly, transmission via fomites and finally, airborne transmission by allowing the volunteers to have either, firstly, direct contact with infants (cuddlers), secondly, touching potential fomites (touchers) or finally, sitting next to the infant (sitters). Volunteers in the group of the cuddlers and touchers but not the sitters became infected, suggesting that direct contact and droplet transmission were the probable routes for efficient infection of the volunteers and that transmission via aerosols was less likely [31]. Another study on the transmission via fomites showed that RSV could be recovered from countertops for several hours, but only for several minutes from absorptive surfaces such as paper tissue and skin  $[32^{\bullet\bullet}]$ . Later on, in the late 1990s, Aintablian et al. detected RSV RNA in the air up to 7 m away from a patient's head [33]. In spite of that, since virus infectivity could not be demonstrated, potential airborne transmission of RSV has been considered negligible and transmission of RSV was thought to occur mainly through contact and droplet transmission. However, in a recent study authors were able to collect aerosols that contained viable virus from the air around RSV infected children [34<sup>••</sup>]. Although the detection of viable virus in the air is by itself not enough to confirm aerosol transmission, the general presumption that RSV exclusively transmits via droplets should be reconsidered and explored further.

### Rhinovirus

Extensive human rhinovirus transmission experiments have not led to a widely-accepted view on the transmission route  $[35-37,38^{\bullet},39^{\bullet},40]$ . Inhalation of aerosols  $(0.2-3 \mu m)$  resulted in efficient rhinovirus infection [41], but little to no infectious rhinovirus could be demonstrated in sneezes and coughs as detected by virus titration [42]. Rhinovirus can survive on stainless steel, plastic and skin for a couple of hours [42,43]. Additionally, virus was detected in saliva, occasionally on hands and could be recovered from the skin of recipients after rubbing either a contaminated fomite or hand [42,44]. When rubbing of fomites was followed by auto-

inoculation this resulted in infection of the volunteers [35]. In a three-day rhinovirus experiment with healthy volunteers different exposure modes were used to investigate the rhinovirus transmission route: Firsrtly, smallparticle exposure (separating donor and recipients by wire mesh), secondly, large particle exposure (encouraging contact, coughing and sneezing while wearing gloves) and finally, direct contact exposure (hand contact followed by self-inoculation). From the results it was concluded that direct contact was the main transmission route [36]. Furthermore, rhinovirus RNA was detected in offices by air sampling studies and subsequent sequencing resulted in a matched air-mucus pair [37]. In a miniature field trail, experimentally infected donors with severe colds participated in a card game with susceptible recipients for  $\sim 12$  hours  $[38^{\bullet\bullet}, 39^{\bullet\bullet}, 40]$ . A restraining device, preventing touching of the head and face, was used in the aerosol condition and heavily contaminated cards and exaggerated hand-to-face movements in the fomite condition. In these experiments aerosol transmission was suggested [40].

In general, transmission rates and exposure time varied between studies, which may contribute to the different routes of transmission that were observed. Therefore, the donor-hours of exposure was determined using donors with severe rhinovirus infections. At 200 hours of exposure to donors, transmission had occurred to 50% of the susceptible recipients, though the transmission route itself was not investigated [38<sup>••</sup>].

#### Influenza A virus

Due to the severity of the yearly influenza epidemics and the potential of zoonotic influenza A viruses to cause severe outbreaks, there have been many studies on influenza A virus transmission among humans. Different kinds of studies, such as air sampling and intervention studies, as well as human challenge studies have been conducted. In addition, transmission events have been described extensively after outbreaks in aircrafts, households and hospital settings. However, until today, results on the relative importance of droplet and aerosol transmission of influenza viruses stay inconclusive and hence, there are many reviews intensively discussing this issue [10,45–50].

Already in the mid-1900s human challenge models were used to assess the transmission route of influenza virus [51°,52–54]. It was shown that illness outcome is dependent on the inoculation route and tends to be milder in intranasally infected volunteers in comparison to inoculation through inhalation [52,53]. Furthermore, illness seemed to be milder in experimentally infected volunteers than in naturally infected individuals [51°]. Increasing numbers of studies focused on the detection and quantification of influenza viruses contained in droplets and aerosols expelled into the air through breathing, sneezing and coughing of infected individuals [9,55–56,57°,58–61]. Influenza virus RNA was detected in the air up to 3.7 m away from patients with the majority of viral RNA contained in aerosols ( $<5 \mu$ m) [59]. The presence of virus in aerosols could indicate potential airborne transmission, although many studies only quantified the amount of viral RNA [55,57°,61]. A few studies quantified viable virus, although this was only recovered from a minority of samples [9,58,59].

#### Coronavirus

In humans, alpha (229E and NL63) and beta coronaviruses (OC43, HKU1, SARS and MERS) are associated with respiratory disease [62,63]. Alpha coronaviruses have a high attack rate early in life and spread rapidly during outbreaks, indicating efficient human to human transmission [63]. Furthermore, samples obtained from staff and patients of a neonatal and pediatric intensive care unit showed a high incidence of human coronaviruses HCoV-229E and HCoV-OC43, suggesting staff-to-patient and patient-to-staff transmission [64]. Unfortunately, there is very little data to corroborate on the HCoV-229E, HCoV-NL63 and HCoV-OC43 transmission routes. HCoV-OC43, HCoV-229E and HCoV-NL63 infectivity was lost between 0 and 72 hours on non-absorptive surfaces, although it can survive several days in medium or PBS [65-67]. Aerosolized HCoV-229E had a half-life of 67 hours in a rotating steel drum (at 20 °C and 50% relative humidity) [68]. SARS-CoV and MERS-CoV appeared to have an unusual capacity to survive on dry surfaces as compared to HCoV-229E, HCoV-OC43, and HCoV-NL63 [69,70].

The SARS outbreak was primarily linked to healthcare settings, with  $\geq 49\%$  of the cases linked to hospitals [71], most probably caused by aerosol-generating procedures on severely ill patients [72,73]. Aerosol-generating procedures like intubation, the use of continuous positivepressure ventilation and drug delivery via nebulizers are likely to produce 'fine infectious droplets', which travel further than droplets from coughs [74]. Additionally, superspreading events contributed to the dispersion of the SARS outbreak [73,75–77], particularly in the Hotel Metropole and the Prince of Wales Hospital in Hong Kong [76]. Moreover, a link with transmission to healthcare workers was observed when they were in close proximity (<1 m) to an index patient, suggesting direct contact or droplet transmission [73,78°,79°]. Air samples and swabs from frequently touched surfaces in a room occupied by a SARS patient tested positive by PCR, although no virus could be cultured from these samples [80]. In the Amoy gardens outbreak fecal droplet transmission was suggested [81,82].

To date, there is little data on the human-to-human MERS-CoV transmission route [83]. MERS-CoV remained stable on non-absorptive for 8 up to 48 hours and for 10 min at  $20 \,^{\circ}$ C and 40% relative humidity in

aerosols [84]. MERS-CoV outbreaks in humans are, like those with SARS-CoV, primarily linked to healthcare settings, with a link to hospitals in  $\geq$ 31% of the cases [71,85,86] and healthcare associated human-to-human transmission was observed [87,88]. Superspreader events were shown to play an important role in nosocomial outbreaks [71,89]. Virus was isolated from environmental samples in hospital rooms, suggesting direct contact or fomite transmission. Moreover, the airborne potential of MERS was investigated by air sample analysis [90,91°]. Viral RNA was detected on the inlet of air ventilation equipment [90] and virus was isolated from air samples and surfaces from inaccessible areas like the ventilator exit, implicating potential aerosol transmission [91°].

## Adenovirus

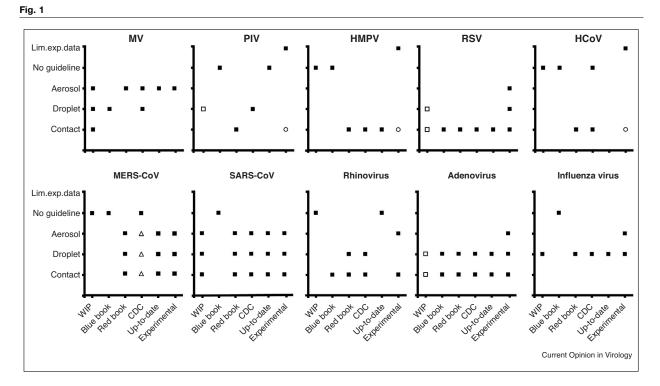
Human adenoviruses can cause respiratory disease (mainly type 1-5, 7, 14 and 21) [92,93], conjunctivitis or infantile gastroenteritis (type 40 and 41) [94]. They are a common cause of respiratory illness and pneumonia in children [95,96], whereas infections are generally asymptomatic in adults [92]. Adenoviruses cause nosocomial outbreaks, especially in pediatric care facilities, where they spread rapidly [95,97,98]. Moreover, adenovirus type 4 and 7 are responsible for large outbreaks of acute respiratory disease, especially in crowded conditions. This is illustrated by, for example, outbreaks among military recruits for which airborne spread was suggested [92,94,99]. It is difficult to eliminate adenovirus from skin, fomites and environmental surfaces [100]. An outbreak in a mental care facility was probably enhanced by spending the day mainly in a crowded room while sharing cigarettes and soda cans, suggesting indirect fomite spread [101]. In a study published in 1966, experimental infections with adenovirus administered as aerosols (0.3- $2.5 \,\mu\text{m}$ ) or droplets (15  $\mu$ m) to healthy, male inmates, resulted in infection of all volunteers, although the resulting illness resembled a natural infection only in the aerosol group [102]. During a military training period, increased numbers of adenovirus infections occurred over time, which correlated with an increased detection of PCR-positive air filters. Additionally, a correlation between disease and the extent of ventilation was observed, with more ventilation resulting in fewer disease cases [103<sup>•</sup>]. In a more recent study in military recruits, positive viral DNA samples were mainly obtained from pillows, lockers and rifles, although adenovirus DNA was also detected in air samples. No consistent correlation between increased positive environmental samples and disease was observed [104].

# Discussion

Studies on the transmission routes of respiratory viruses have been performed since the beginning of the 20th century [105]. Despite this, the relative importance of transmission routes of respiratory viruses is still unclear, depending on the heterogeneity of many factors like the environment (e.g. temperature and humidity), pathogen and host [5,19]. Differences in virus shedding between individuals can contribute to the transmissibility rate, especially in the case of superspreaders [75,106]. In addition, the SARS-CoV outbreak highlighted the impact of aerosol-generating procedures on the increased risk of human-to-human transmission [74,107], demonstrating that for these procedures additional containment measures are necessary.

Inter-human transmission has been studied under many different (experimental) conditions. A summary of the advantages and disadvantages of the different study designs (Table 3) highlights the difficulty of human transmission experiments. As a consequence, contrasting results have been obtained for many viruses. This is also reflected in Table 2, summarizing the experimental data on inter-human transmission. Besides the difficulty of performing studies under well-controlled conditions, another key issue is that often (attenuated) laboratory strains are studied in healthy adults, which does not reflect the natural circumstances and target group and hence influence the outcome of the studies.

Respiratory viruses are an important cause of nosocomial infections, especially in children. Therefore, we consulted the guidelines on infection prevention from National [108], European [109], American [3,110] and International [111]) organizations for their information on transmission routes (Table 2) and associated isolation guidelines (Figure 1). Unfortunately, terms and definitions of respiratory transmission routes and isolation guidelines are not always used in a uniform way, leaving room for personal interpretation. But more importantly, information on the transmission route does not always reflect the isolation guidelines (e.g. for PIV and rhinovirus, Figure 1). As a proxy for transmission route, virus stability is often referred to in the guidelines, however, this can only imply a role for indirect contact transmission but is by no means conclusive on the transmission route. In hospital settings, prevention of contact transmission is generally implemented in standard infection prevention



Isolation guidelines for respiratory virus infections in comparison to experimental evidence on transmission routes. Isolation guidelines for all respiratory viruses discussed in this review from National (Working Group Infection Prevention (WIP) [108], from the Netherlands National Institute for Public Health and the Environment (RIVM), European ('The Blue Book' [109]), American ('The Red Book' [110] and the Centers for Disease Control (CDC) [3]) and International (UpToDate [111]) organizations are shown on the X-axis, together with the experimental evidence on transmission routes (Table 2). The categories on the Y-axis are the different transmission routes (contact, droplet or aerosol), the absence of guidelines for infection prevention ('No guideline'), or the limited availability of experimental data ('Lim. exp. data'). The information shown for influenza virus reflects the guidelines on seasonal influenza virus. Closed squares ( $\blacksquare$ ): isolation guidelines for the respective respiratory virus. Open squares ( $\square$ ): guidelines are only for children  $\leq 6$  years old. Open circles ( $\diamond$ ) tata from stability experiments only. Open triangles ( $\triangle$ ): specific CDC guidelines for Healthcare Professionals [115] (not the isolation guideline [3] used in this review).

precautions such as strict hand hygiene and cough etiquette. It is important to note differences in isolation guidelines between different organizations and the lack of correlation to scientific data. The variation in described transmission routes and associated isolation guidelines among the different organizations underscores the lack of convincing data.

Well-designed human infection studies could be employed to investigate the role of transmission routes of respiratory viruses among humans [112\*\*]. However, since human transmission experiments are very challenging, animal transmission models can provide an attractive alternative and should be explored and developed for all respiratory viruses. In such experiments, the influence of environmental factors on transmission routes can also be investigated [113]. However, before extrapolating experimentally generated data to humans, it is important to understand the limitations of these models, and appreciate the heterogeneity of experimental setups employed in laboratories [114]. Furthermore, quantitative data such as viral load in the air can be obtained by air sampling methods in various environments, such as hospital settings. Air sampling of viruses is an increasingly used technology in animal and human experiments. However, whereas most studies rely on the detection of viral genome copies, viability assays such as plaque assays or virus titration should be included to gain information on virus infectivity.

Ultimately, the knowledge gap on inter-human transmission should be filled by developing and performing stateof-the art experiments in a natural setting. Combined with animal transmission models and air sampling in different (health care and experimental) settings, these data should result in a thorough scientific understanding of the inter-human transmission routes of respiratory viruses. Eventually, this knowledge will help with an evidence-based risk assessment of the different transmission routes to improve existing infection prevention strategies.

#### Acknowledgements

We thank Dr. Rik de Swart, Dr. Bart Haagmans, Dr. Arno Andeweg, and Dr. Sabrina Schreiner for helpful discussions. JK and SH are supported by an NWO VIDI grant (contract number 91715372), and MS, RF and SH by NIAID/NIH contract HHSN272201400008C. PF receives funding from the EU FP7 project PREPARE (grant number 602525). The sponsors had no role in the collection, analysis and interpretation of data, in the writing of the report, and in the decision to submit the article for publication.

#### References and recommended reading

Papers of particular interest, published within the period of review, have been highlighted as:

- of special interest
- •• of outstanding interest
- 1. WHO. Available from: http://www.who.int/en/ [cited 2017 28.08.17].

- Ching PH, Li K, Pessoa-Silva Y, Seto CL, Wang WTKF: Infection prevention and control of epidemic and pandemic-prone acute respiratory diseases in health care: WHO interim guidelines. Geneva: WHO; 2007, 90.
- Siegel JD et al.: 2007 Guideline for isolation precautions: preventing transmission of infectious agents in health care settings. Am J Infect Control 2007, 35(Suppl 2):S65-S164.
- 4. Gratton J et al.: The role of particle size in aerosolised pathogen transmission: a review. J Infect 2011, 62:1-13.
- 5. CDC. 2017. Available from: https://www.cdc.gov/ [cited 28.08.17].
- Weber TP, Stilianakis NI: Inactivation of influenza A viruses in the environment and modes of transmission: a critical review. *J Infect* 2008, 57:361-373.
- Milton DK et al.: Influenza virus aerosols in human exhaled breath: particle size, culturability, and effect of surgical masks. PLoS Pathog 2013, 9:e1003205.
- Tellier R: Aerosol transmission of influenza A virus: a review of new studies. J R Soc Interface 2009, 6(Suppl 6):S783-S790.
- Pica N, Bouvier NM: Environmental factors affecting the transmission of respiratory viruses. Curr Opin Virol 2012, 2:90-95.
- Fernstrom A, Goldblatt M: Aerobiology and its role in the transmission of infectious diseases. J Pathog 2013, 2013:493960.
- 11. Herfst S et al.: Drivers of airborne human-to-human pathogen transmission. Curr Opin Virol 2017, 22:22-29.
- 12. Moss WJ, Griffin DE: Measles. Lancet 2012, 379:153-164.
- WHO: Measles. 2017. Available from: http://www.who.int/ith/ diseases/measles/en/ [cited 07.08.17].
- Bloch AB et al.: Measles outbreak in a pediatric practice:
   airborne transmission in an office setting. Pediatrics 1985, 75:676-683

Detailed retrospective analysis including airflow studies of a measles outbreak in a pediatric practice.

- Chen RT et al.: An explosive point-source measles outbreak in a highly vaccinated population, modes of transmission and risk factors for disease. Am J Epidemiol 1989, 129:173-182.
- Ehresmann KR et al.: An outbreak of measles at an international sporting event with airborne transmission in a domed stadium. J Infect Dis 1995, 171:679-683.
- Riley EC, Murphy G, Riley RL: Airborne spread of measles in a suburban elementary school. Am J Epidemiol 1978, 107:421-432.
- 18. Remington PL et al.: Airborne transmission of measles in a
   physician's office. J Am Med Assoc 1985, 253:1574-1577.
   Detailed retrospective analysis including airflow studies and a mathematical airborne transmission model of a measles outbreak in a physician's office.
- Stein RA: Super-spreaders in infectious diseases. Int J Infect Dis 2011, 15:e510-e513.
- 20. Lau MS et al.: Spatial and temporal dynamics of superspreading events in the 2014-2015 West Africa Ebola epidemic. Proc Natl Acad Sci U S A 2017, 114:2337-2342.
- Lloyd-Smith JO, Kopp SS, Getz PEWM: Superspreading and the effect of individual variation on disease emergence. *Nature* 2005, 438:355-359.
- Christensen PE et al.: An epidemic of measles in southern Greenland, 1951; measles in virgin soil. II. The epidemic proper. Acta Med Scand 1953, 144:430-449.
- CDC: Human Parainfluenza Viruses (HPIVs). 2015. Available from: https://www.cdc.gov/parainfluenza/about/transmission.html [cited 26.07.17].
- Henrickson KJ: Parainfluenza viruses. Clin Microbiol Rev 2003, 16:242-264.

- Karron RA, Collins PL: Parainfluenza viruses. In *Fields Virology*. Edited by Wolters Kluwer LWW. Lippincott Williams & Wilkins: Wolters Kluwer; 2013:996-1023.
- Brady MT, Evans J, Cuartas J: Survival and disinfection of parainfluenza viruses on environmental surfaces. Am J Infect Control 1990, 18:18-23.
- Ansari SA et al.: Potential role of hands in the spread of respiratory viral infections: studies with human parainfluenza virus 3 and rhinovirus 14. J Clin Microbiol 1991, 29:2115-2119.
- Miller WS, Artenstein MS: Aerosol stability of three acute respiratory disease viruses. Proc Soc Exp Biol Med 1967, 125:222-227.
- McLean DM, Bannatyne RM, Givan KF: Myxovirus dissemination by air. Can Med Assoc J 1967, 96:1449-1453.
- Tollefson SJ, Cox RG, Williams JV: Studies of culture conditions and environmental stability of human metapneumovirus. *Virus Res* 2010, 151:54-59.
- Hall CB, Douglas RG Jr: Modes of transmission of respiratory syncytial virus. J Pediatr 1981, 99:100-103.
- Hall CB, Douglas RG Jr, Geiman JM: Possible transmission by fomites of respiratory syncytial virus. J Infect Dis 1980, 141:98-102.

Study in which transmission modes of RSV between infants and adults were investigated.

- Aintablian N, Walpita P, Sawyer MH: Detection of Bordetella pertussis and respiratory synctial virus in air samples from hospital rooms. Infect Control Hosp Epidemiol 1998, 19:918-923.
- Kulkarni H et al.: Evidence of respiratory syncytial virus spread
   by aerosol, time to revisit infection control strategies? Am J Respir Crit Care Med 2016, 194:308-316.

Air-sampling study in a children's hospital in which viable RSV was collected from the air.

- Gwaltney JM Jr, Hendley JO: Transmission of experimental rhinovirus infection by contaminated surfaces. Am J Epidemiol 1982, 116:828-833.
- Gwaltney JM Jr, Moskalski PB, Hendley JO: Hand-to-hand transmission of rhinovirus colds. Ann Intern Med 1978, 88:463-467.
- Myatt TA et al.: Detection of airborne rhinovirus and its relation to outdoor air supply in office environments. Am J Respir Crit Care Med 2004, 169:1187-1190.
- Meschievitz CK, Schultz SB, Dick EC: A model for obtaining
   predictable natural transmission of rhinoviruses in human volunteers. J Infect Dis 1984. 150:195-201.

In this study the importance of exposure time in human transmission experiments was demonstrated.

 39. Dick EC *et al.*: Interruption of transmission of rhinovirus colds
 among human volunteers using virucidal paper handkerchiefs. *J Infect Dis* 1986, 153:352-356.

In this study the miniature field trial was used to discriminate between aerosol and indirect contact transmission.

- Dick EC et al.: Aerosol transmission of rhinovirus colds. J Infect Dis 1987, 156:442-448.
- Cate TR et al.: Production of tracheobronchitis in volunteers with rhinovirus in a small-particle aerosol. Am J Epidemiol 1965, 81:95-105.
- Hendley JO, Wenzel RP, Gwaltney JM Jr: Transmission of rhinovirus colds by self-inoculation. N Engl J Med 1973, 288:1361-1364.
- Reed SE: An investigation of the possible transmission of Rhinovirus colds through indirect contact. J Hyg (Lond) 1975, 75:249-258.
- 44. Pancic F, Carpentier DC, Came PE: Role of infectious secretions in the transmission of rhinovirus. *J Clin Microbiol* 1980, **12**:567-571.
- 45. Brankston G et al.: Transmission of influenza A in human beings. Lancet Infect Dis 2007, 7:257-265.

- 46. Lee RV: Transmission of influenza A in human beings. Lancet Infect Dis 2007, 7:761-763 760-1; author reply.
- Lemieux C et al.: Questioning aerosol transmission of influenza. Emerg Infect Dis 2007, 13(1):173-174 author reply 174-5.
- Tellier R: Transmission of influenza A in human beings. Lancet Infect Dis 2007, 7:759-760 author reply 761-3.
- 49. Tellier R: Review of aerosol transmission of influenza A virus. Emerg Infect Dis 2006, **12**:1657-1662.
- Killingley B, Nguyen-Van-Tam J: Routes of influenza transmission. Influenza Other Respir Viruses 2013, 7(Suppl 2):42-51.

51. Little JW *et al.*: Attenuated influenza produced by experimental
intranasal inoculation. J Med Virol 1979, 3:177-188.

Authors compared natural and experimentally induced influenza infections by assessing clinical illness and pulmonary function.

- Alford RH et al.: Human influenza resulting from aerosol inhalation. Proc Soc Exp Biol Med 1966, 122:800-804.
- Henle W, Henle G et al.: Experimental exposure of human subjects to viruses of influenza. J Immunol 1946, 52:145-165.
- Smorodintseff AAT, Drobyshevskaya MD, Korovin AI, Osetroff AAAI: Investigation on volunteers infected with the influenza virus. Am J Med Sci 1937, 194:159-170.
- Bischoff WE et al.: Exposure to influenza virus aerosols during routine patient care. J Infect Dis 2013, 207:1037-1046.
- Fabian P et al.: Influenza virus in human exhaled breath: an observational study. PLoS ONE 2008, 3:e2691.
- 57. Lindsley WG et al.: Viable influenza A virus in airborne particles
   expelled during coughs versus exhalations. Influenza Other Respir Viruses 2016, 10:404-413.

Authors compared the generation of aerosols laden with viable influenza virus during coughing and exhalation of infected individuals.

- Lindsley WG et al.: Measurements of airborne influenza virus in aerosol particles from human coughs. PLoS ONE 2010, 5: e15100.
- Lednicky JA, Loeb JC: Detection and isolation of airborne influenza A H3N2 virus using a sioutas personal cascade impactor sampler. Influenza Res Treat 2013, 2013:656825.
- Blachere FM et al.: Measurement of airborne influenza virus in a hospital emergency department. Clin Infect Dis 2009, 48: 438-440.
- Lindsley WG et al.: Distribution of airborne influenza virus and respiratory syncytial virus in an urgent care medical clinic. Clin Infect Dis 2010, 50:693-698.
- Adams MJ et al.: Changes to taxonomy and the International Code of Virus Classification and Nomenclature ratified by the International Committee on Taxonomy of Viruses (2017). Arch Virol 2017.
- Masters PS, Perlman S: Coronaviridae. In Fields virology. Edited by Knipe HDMPM. Wolters Kluwer, Lippcott Williams & Wilkins; 2007:825-858.
- Gagneur A et al.: Coronavirus-related nosocomial viral respiratory infections in a neonatal and paediatric intensive care unit: a prospective study. J Hosp Infect 2002, 51:59-64.
- Sizun J, Yu MW, Talbot PJ: Survival of human coronaviruses 229E and OC43 in suspension and after drying on surfaces: a possible source of hospital-acquired infections. J Hosp Infect 2000, 46:55-60.
- 66. Muller A *et al.*: Stability of human metapneumovirus and human coronavirus NL63 on medical instruments and in the patient environment. *J Hosp Infect* 2008, 69:406-408.
- Rabenau HF et al.: Stability and inactivation of SARS coronavirus. Med Microbiol Immunol (Berl) 2005, 194:1-6.
- 68. Ijaz MK et al.: Survival characteristics of airborne human coronavirus 229E. J Gen Virol 1985, 66:2743-2748.

- 150 Emerging viruses: intraspecies transmission
- 69. Otter JA et al.: Transmission of SARS and MERS coronaviruses and influenza virus in healthcare settings: the possible role of dry surface contamination. J Hosp Infect 2016, 92:235-250
- 70. Duan SM et al.: Stability of SARS coronavirus in human specimens and environment and its sensitivity to heating and UV irradiation. Biomed Environ Sci 2003. 16:246-255
- 71. Chowell G et al.: Transmission characteristics of MERS and SARS in the healthcare setting: a comparative study. BMC Med 2015. 13:210.
- Lee N et al.: A major outbreak of severe acute respiratory syndrome in Hong Kong. N Engl J Med 2003, 348:1986-1994.
- 73. Ofner M et al.: Cluster of severe acute respiratory syndrome cases among protected health-care workers - Toronto, Canada, April (Reprinted from MMWR, vol. 52, pg 433-436, 2003). J Am Med Assoc 2003. 289:2788-2789
- 74. Gamage B et al.: Protecting health care workers from SARS and other respiratory pathogens: a review of the infection control literature. Am J Infect Control 2005, 33:114-121.
- 75. Wong G et al.: MERS, SARS, and Ebola: the role of superspreaders in infectious disease. Cell Host Microbe 2015, 18:398-401
- 76. Braden CR et al.: Progress in global surveillance and response capacity 10 years after severe acute respiratory syndrome. Emerg Infect Dis 2013, 19:864-869.
- 77. Shen Z et al.: Superspreading SARS events, Beijing, 2003. Emerg Infect Dis 2004, 10:256-260.
- 78. Wong TW et al.: Cluster of SARS among medical students exposed to single patient, Hong Kong. Emerg Infect Dis 2004, 10.269-276

Authors demonstrate SARS-CoV transmission in healthcare workers and a link with proximity to the index case.

Varia M et al.: Investigation of a nosocomial outbreak of severe acute respiratory syndrome (SARS) in Toronto, Canada. CMAJ 79. 2003. 169:285-292

Authors describe the risk of SARS-CoV transmission regarding distance and aerosol-generating procedures during a nosocomial outbrea

- 80. Booth TF et al.: Detection of airborne severe acute respiratory syndrome (SARS) coronavirus and environmental contamination in SARS outbreak units. J Infect Dis 2005, 191:1472-1477
- 81. Poutanen SM, McGeer AJ: Transmission and control of SARS. Curr Infect Dis Rep 2004, 6:220-227.
- 82. Yu IT et al.: Evidence of airborne transmission of the severe acute respiratory syndrome virus. N Engl J Med 2004, 350:1731-1739
- 83. Shapiro M et al.: Middle East respiratory syndrome coronavirus: review of the current situation in the world. Disaster Mil Med 2016, 2:9.
- 84. van Doremalen N, Bushmaker T, Munster VJ: Stability of Middle East respiratory syndrome coronavirus (MERS-CoV) under different environmental conditions. Euro Surveill 2013, 18.
- Hunter JC et al.: Transmission of middle east respiratory 85. syndrome coronavirus infections in healthcare settings, Abu Dhabi. Emerg Infect Dis 2016, 22:647-656.
- Oboho IK et al.: 2014 MERS-CoV outbreak in Jeddah a link to health care facilities. N Engl J Med 2015, 372:846-854. 86.
- 87. Assiri A et al.: Hospital outbreak of Middle East respiratory syndrome coronavirus. N Engl J Med 2013, 369:407-416.
- 88. Guery B et al.: Clinical features and viral diagnosis of two cases of infection with Middle East Respiratory Syndrome coronavirus: a report of nosocomial transmission. Lancet 2013, 381:2265-2272.
- 89. Oh MD et al.: Middle east respiratory syndrome coronavirus Superspreading event involving 81 persons, Korea. J Korean Med Sci 2015, 30:1701-1705.

- 90. Bin SY et al.: Environmental contamination and viral shedding in MERS patients during MERS-CoV outbreak in South Korea. Clin Infect Dis 2016. 62:755-760.
- 91. Kim SH *et al.*: Extensive viable middle east respiratory syndrome (MERS) coronavirus contamination in air and surrounding environment in MERS isolation wards. *Clin Infect Dis* 2016, **63**:363-369.

First time that MERS-CoV was demonstrated in air samples from outbreak units. Virus was also detected on fomites and environmental surfaces

- 92. Musher DM: How contagious are common respiratory tract infections? N Engl J Med 2003, 348:1256-1266
- 93. Scott MK et al.: Human adenovirus associated with severe respiratory infection, Oregon, USA, 2013-2014. Emerg Infect Dis 2016. 22:1044-1051.
- Berk AJ: Adenoviridae. In *Fields virology*. Edited by Knipe HDDM/PI/VI31Lippincott Williams & Wilkins: Wolters Kluwer; 2013:
- 95. Hatherill M et al.: Evolution of an adenovirus outbreak in a multidisciplinary children's hospital. J Paediatr Child Health 2004, 40:449-454
- 96. Palomino MA, Larranaga C, Avendano LF: Hospital-acquired adenovirus 7h infantile respiratory infection in Chile. Pediatr Infect Dis J 2000, 19:527-531.
- James L et al.: Outbreak of human adenovirus type 3 infection 97. in a pediatric long-term care facility - Illinois, 2005. Clin Infect Dis 2007. 45:416-420.
- Porter JD et al.: Outbreak of adenoviral infections in a long-98. term paediatric facility, New Jersey, 1986/87. J Hosp Infect 1991, 18:201-210.
- 99. Hilleman MR: Epidemiology of adenovirus respiratory infections in military recruit populations. Ann N Y Acad Sci 1957 67:262-272
- 100. Forgie S, Marrie TJ: Healthcare-associated atypical pneumonia. Semin Respir Crit Care Med 2009, 30:67-85
- 101. Klinger JR et al.: Multiple cases of life-threatening adenovirus pneumonia in a mental health care center. Am J Respir Crit Care Med 1998. 157:645-649.
- 102. Couch RB et al.: Aerosol-induced adenoviral illness resembling the naturally occurring illness in military recruits. Am Rev Respir Dis 1966. 93:529-535.
- 103. Echavarria M et al.: Detection of adenoviruses (AdV) in culture negative environmental samples by PCR during an AdV-associated respiratory disease outbreak. J Clin Microbiol 2000, 38:2982-2984

Authors show a link between adenovirus DNA in air filters and adenovirusrelated hospitalizations.

- 104. Russell KL et al.: Transmission dynamics and prospective environmental sampling of adenovirus in a military recruit setting. J Infect Dis 2006, 194:877-885.
- 105. Thomson FH, Price C: The aerial conveyance of infection. Lancet 1914:1669-1673
- 106. Leavy O: Infectious disease: the tolerance of superspreaders. Nat Řev Immunol 2014, 14:776-777.
- 107. Christian MD et al.: Possible SARS coronavirus transmission during cardiopulmonary resuscitation. Emerg Infect Dis 2004, 10:287-293
- 108. WIP: W.i.p. Ziekenhuizen: indicatie voor isolatie. 2006-2013. Available from: http://www.rivm.nl/dsresource?objectid=6642621a-52da-4cd0-96c7-58bea053b380&type=org&disposition=inline [cited 01.11.17].
- 109. Sharland M, Butler K, Giaquinto C, Finn A, Esposito S, Galanakis M: Manual of childhood infections: the blue book. edn 4. Oxford University Press; 2016.
- 110. Pickering LK, Baker CJ, Kimberlin DW, Long SH: Red book: 2012 report of the committee on infectious diseases. edn 29. Elk Grove Vilage: American Academy of Pediatrics; 2012.

- 111. UpToDate. 2017. Available from: https://www.uptodate.com/ [cited 20.10.17].
- 112. Killingley B et al.: Use of a human influenza challenge model to
   assess person-to-person transmission: proof-of-concept study. J Infect Dis 2012, 205:35-43.

A human proof-of-concept study, in which authors assessed infectivity rate of experimentally inoculated volunteers and secondary attack rate among susceptible individuals.

- 113. Lowen AC et al.: Influenza virus transmission is dependent on relative humidity and temperature. PLoS Pathog 2007, 3:1470-1476.
- 114. Belser JA et al.: Complexities in ferret influenza virus pathogenesis and transmission models. Microbiol Mol Biol Rev 2016, 80:733-744.
- CDC: Interim infection prevention and control recommendations for hospitalized patients with Middle East Respiratory Syndrome Coronavirus (MERS-CoV). 2015. Available from: https://www.cdc. gov/coronavirus/mers/infection-prevention-control.html [cited 13.11.17].
- 116. Knight V: Viruses as agents of airborne contagion. Ann NY Acad Sci 1980, 353:147-156.
- 117. Seto WH et al.: Effectiveness of precautions against droplets and contact in prevention of nosocomial transmission of severe acute respiratory syndrome (SARS). Lancet 2003, 361:1519-1520.
- Olsen SJ et al.: Transmission of the severe acute respiratory syndrome on aircraft. N Engl J Med 2003, 349:2416-2422.
- 119. D'Alessio DJ et al.: Transmission of experimental rhinovirus colds in volunteer married couples. J Infect Dis 1976, 133:28-36.
- 120. Aditama TY et al.: Avian influenza H5N1 transmission in households, Indonesia. PLoS ONE 2012, 7:e29971.
- 121. Hall CB et al.: Respiratory syncytial virus infections within families. N Engl J Med 1976, 294:414-419.
- 122. Chu HY et al.: Nosocomial transmission of respiratory syncytial virus in an outpatient cancer center. Biol Blood Marrow Transpl 2014, 20:844-851.
- 123. Lau JT et al.: Probable secondary infections in households of SARS patients in Hong Kong. Emerg Infect Dis 2004, 10:235-243.
- 124. Coleman KP, Markey PG: Measles transmission in immunized and partially immunized air travellers. *Epidemiol Infect* 2010, 138:1012-1015.
- 125. Amornkul PN et al.: Low risk of measles transmission after exposure on an international airline flight. J Infect Dis 2004, 189 (Suppl 1):S81-S85.
- 126. Neatherlin J et al.: Influenza A(H1N1)pdm09 during air travel. Travel Med Infect Dis 2013, 11:110-118.

- 127. Shankar AG et al.: Contact tracing for influenza A(H1N1)pdm09 virus-infected passenger on international flight. Emerg Infect Dis 2014, 20:118-120.
- 128. Cowling BJ et al.: Facemasks and hand hygiene to prevent influenza transmission in households: a cluster randomized trial. Ann Intern Med 2009, 151:437-446.
- 129. Simmerman JM et al.: Findings from a household randomized controlled trial of hand washing and face masks to reduce influenza transmission in Bangkok, Thailand. Influenza Other Respir Viruses 2011, 5:256-267.
- 130. Allison MA et al.: Feasibility of elementary school children's use of hand gel and facemasks during influenza season. Influenza Other Respir Viruses 2010, 4:223-229.
- 131. Hayden GF, Hendley JO, Gwaltney JM Jr: The effect of placebo and virucidal paper handkerchiefs on viral contamination of the hand and transmission of experimental rhinoviral infection. J Infect Dis 1985, 152:403-407.
- 132. Kaiser L et al.: Short-term treatment with zanamivir to prevent influenza: results of a placebo-controlled study. Clin Infect Dis 2000, 30:587-589.
- 133. Bischoff WE et al.: Detection of measles virus RNA in air and surface specimens in a hospital setting. J Infect Dis 2016, 213:600-603.
- 134. Lidwell OM, Towers AG: Protection from microbial contamination in a room ventilated by a uni-directional air flow. J Hyg (Lond) 1969, 67:95-106.
- 135. Normile D: The Metropole, superspreaders, and other mysteries. Science 2013, 339:1272-1273.
- 136. Beggs CB, Shepherd SJ, Kerr KG: Potential for airborne transmission of infection in the waiting areas of healthcare premises: stochastic analysis using a Monte Carlo model. *BMC Infect Dis* 2010, 10:247.
- 137. Nicas M, Jones RM: Relative contributions of four exposure pathways to influenza infection risk. *Risk Anal* 2009, 29:1292-1303.
- 138. Wagner BG, Coburn BJ, Blower S: Calculating the potential for within-flight transmission of influenza A (H1N1). BMC Med 2009, 7:81.
- 139. Nicas M, Best D: A study quantifying the hand-to-face contact rate and its potential application to predicting respiratory tract infection. J Occup Environ Hyg 2008, 5:347-352.
- 140. Tang JW et al.: Factors involved in the aerosol transmission of infection and control of ventilation in healthcare premises. J Hosp Infect 2006, 64:100-114.
- 141. Chowell G et al.: Synthesizing data and models for the spread of MERS-CoV. 2013, key role of index cases and hospital transmission. Epidemics 2014, 9:40-51.